

Table 12: Summary of dietary exposure to E 171 from its use as a food additive in the maximum level exposure assessment scenario and in the refined exposure scenarios, in six population groups (minimum–maximum across the dietary surveys in mg/kg bw per day)

	Infants (12 weeks-11 months)	Toddlers (12–35 months)	Children (3–9 years)	Adolescents (10–17 years)	Adults (18– 64 years)	The elderly (≥ 65 years)	
Maximum level exposure assessment scenario							
Mean95th percentile	0.06–3.6 0.2–15.8	0.9–12.8 2.9–31.4	1.9–11.5 5.9–31.3	1.3–6.2 4.0–18.6	0.7–6.7 2.4–15.9	0.4–4.9 1.9–12.7	
Refined exposure assessment scenarios							
Brand-loyal scenario							
Mean95th percentile	0.05–3.5 0.1–14.3	0.8–10.0 2.6–28.0	1.7–9.7 5.2–25.4	1.1–5.0 3.3–14.9	0.6–5.5 2.0–13.1	0.4–4.2 1.7–10.4	
Non-brand-loyal scenario							
Mean95th percentile	0.03–2.9 0.1–9.9	0.6–6.0 1.9–27.5	0.9–6.9 2.5–23.7	0.6–3.6 1.6–13.2	0.3–3.8 1.2–9.5	0.2–2.8 0.9–7.1	

bw: body weight.

In the *maximum level exposure assessment scenario*, mean exposure to E 171 from its use as a food additive ranged from 0.06 mg/kg bw per day in infants to 12.8 mg/kg bw per day in toddlers. The 95th percentile of exposure ranged from 0.2 mg/kg bw per day in infants to 31.4 mg/kg bw per day in toddlers.

In the *brand-loyal refined exposure assessment scenario*, mean exposure to E 171 from its use as a food additive ranged from 0.05 mg/kg bw per day in infants to 10.0 mg/kg bw per day in toddlers. The 95th percentile of exposure ranged from 0.1 mg/kg bw per day in infants to 28.0 mg/kg bw per day in toddlers. In the *non-brand-loyal scenario*, mean exposure ranged from 0.03 mg/kg bw per day in infants to 6.9 mg/kg bw per day in children. The 95th percentile of exposure ranged from 0.1 mg/kg bw per day in infants to 27.5 mg/kg bw per day in toddlers.

It can be noted that for infants, toddlers, children and adolescents, the maximum of the range of dietary exposure calculated is higher than in the 2016 EFSA ANS opinion. This is due to one dietary survey which was recently included in the EFSA Comprehensive database.

For the *food supplements consumers only* (results reported in Appendix T), mean exposure to E 171 from its use as a food additive ranged from 0.8 mg/kg bw per day for adults to 11.7 mg/kg bw per day for children. The 95th percentile ranged from 3.1 mg/kg bw per day for the elderly to 41 mg/kg bw per day for children.

The main food categories contributing to the exposure to E 171 are presented in Appendix U. In the non-brand-loyal exposure assessment scenario, the main contributing food categories for infants, toddlers and adolescents were fine bakery wares, soups and broths and sauces; they were soups and broths, sauces and salads and savoury-based sandwich spreads for children, adults and the elderly. Processed nuts were also a main contributing food category for adults and the elderly (Appendix U).

An exposure estimate for E 171 considering data from scientific publications (Weir et al., 2012; Taboada-López et al., 2019) was also performed. The six additional food categories considered were unripened cheeses (FC 01.7.1), processed cheeses (FC 01.7.5), breakfast cereals (FC 06.3), rice (FC 06.7), surimi (FC 09.2) and snacks (FC 15.1). Levels of E 171 for these foods were based on analytical data and were low compared to use levels reported for the other food categories. It is uncertain if products from these additional food categories are available on the European market, and therefore, the relevance of the data is unclear. However, even taking these six additional food categories into consideration resulted in only a small increase in the calculated dietary exposure to E 171 (up to an additional 0.01 mg/kg bw per day for some populations groups) (Appendix V). Some publications reported levels of E 171 in foods where E 171 is not authorised to be added in the EU (e.g. raw cows milk, fresh milk, long-life milk, plain yoghurt (Rompelberg et al., 2016); chocolate products, ripened cheese, (Weir et al., 2012) and seafood products (Yin et al., 2017). As these are not authorised uses in the EU, these levels were not considered in the above assessment.



Uncertainty analysis

In accordance with the guidance provided in the EFSA opinion related to uncertainties in dietary exposure assessment (EFSA, 2007), the following sources of uncertainties have been considered and summarised in Table 13.

Table 13: Qualitative evaluation of influence of uncertainties on the dietary exposure estimate

Sources of uncertainties	Direction ^(a)	
Consumption data: different methodologies/representativeness/underreporting/misreporting/no portion size standard		
Methodology used to estimate high percentiles (95th) long-term (chronic) exposure based on data from food consumption surveys covering only a few days		
Correspondence of reported use levels and analytical data to the food items in the EFSA Comprehensive Database: uncertainties to which types of food the levels refer	+/-	
Uncertainty in possible national differences in use levels of food categories	+/-	
Occurrence data: — use levels considered applicable to all foods within the entire food category, whereas the average percentage of food products in the EU labelled as containing E 171 was only 1% of all the food products in the subcategories in Mintel in which E 171 is listed — not fully representative of foods on the EU market	+/-	
The 16 food categories which were taken into account in the refined exposure assessment scenarios out of all authorised food categories ($N = 48$), corresponded to different percentage, depending on the food categories (32% – 96% of the amount (gram of foods by body weight) of food consumption documented in the EFSA Comprehensive Database)	-	
Food categories included in the exposure assessment: no data for certain food categories which were therefore not considered in the exposure estimates $(n = 32/48)$	_	
Maximum level exposure assessment scenario: — exposure calculations based on the maximum reported use levels (reported use from industries)	+	
Refined exposure assessment scenarios: — exposure calculations based on the maximum or mean levels (reported use from industries)	+/_	

⁽a): +, uncertainty with potential to cause overestimation of exposure; -, uncertainty with potential to cause underestimation of exposure.

E 171 is authorised in four food categories and listed among the food colours of Group II authorised in other 44 food categories (Table 10).

In the current exposure assessment, 16 food categories were considered in the maximum level and in the refined scenarios. These food categories are the main food categories in terms of consumption levels. The Panel calculated that out of the foods authorised to contain E 171 according to Annex II to Regulation (EC) No 1333/2008, 32–96% of the amount of food consumed (by weight) per population group was reported to potentially contain E 171 as a food additive.

The Panel noted that information from the Mintel's GNPD (Appendix R) indicated that 49 out of 86 food subcategories, categorised according to the Mintel's GNPD nomenclature, in which E 171 was labelled were included in the current exposure assessment. These 49 food subcategories represented approximately 95% of the food products labelled with E 171 in the database. Furthermore, the percentage of foods per subcategory labelled to contain E 171 was maximally about 52% in sticks, liquids and sprays (on average, the percentage of food products in the EU labelled as containing E 171 was 1% of all the food products included in the subcategories in Mintel in which E 171 is listed) (Appendix R), while in the assessment, it was assumed that 100% of the foods belonging to an authorised food category contained the food additive.

Given these observations, the Panel considered overall that the uncertainties identified resulted in an overestimation of the exposure to E 171 from its use as a food additive according to Annex II to Regulation No 1333/2008 for all scenarios.



Oral exposure via other sources

E 171 is widely used as an excipient in medicinal products (Section 3). No information on the extent and level of use of E 171 in medicinal products was made available to EFSA and, therefore, its exposure from this use could not be considered. Exposure to TiO_2 via cosmetics (e.g. toothpaste) is not considered in this opinion.

4.4.4. Exposure to TiO₂ NPs from the use of E 171

Number-based particle size distributions of a series of pristine E 171 food additives and of E 171 particles extracted from food products have been reported (EFSA FAF Panel, 2019; Verleysen et al., 2020, 2021; Geiss et al., 2021). In these publications, the size and shape of isolated constituent particles and of constituent particles in aggregates and agglomerates were measured by image analysis of representative transmission electron micrographs using ellipse fitting. The short (a-axis) and long axes (c-axis) of the ellipse fitted to the 2D projection of each constituent particle was measured as a proxy of its minimal and maximal external dimension, respectively. Assuming that the particles are prolate ellipsoids (a axis = b axis < c axis), the volume (V) of each particle was estimated as V = $^4/_3 \cdot \pi \cdot a^2$ c and the mass (M) of each particle was calculated as M = V ϕ , with ϕ = the density of the particles (3.89 g/cm³ for anatase and 4.32 g/cm³ for rutile particles). More information on the analysis of E 171 is available in the report of a project focusing on the development of analytical methodologies that allow identification and characterisation of nanoparticles in food additives in their pristine state and in simple food matrices (Verleysen et al., 2021). Using this methodology, the mass and number percentages of particles in different samples of E 171 were calculated (also Appendix W; reported in Verleysen et al., 2021).

Taking into account the available data (Verleysen et al., 2021), it can be presumed that the mass of constituent particles below 100 nm could be up to 30%, where the mean of the 12 analysed samples is 25%. The Panel noted that different types of E 171 are used in food and the percentage by number of constituent particles below 100 nm can range from 5% (rutile) to around 50% (EFSA FAF Panel, 2019). The use levels of the different types of E 171 are unknown, and therefore, it is not possible to estimate accurately the exposure to nanoparticulate TiO₂ from the use of E 171 (Table 12).

5. Uncertainty considerations

The Panel, after evaluating the scientific evidence available, has identified uncertainties related to the following points:

- The size distribution of the particles in marketed E 171 that consumers are exposed to, related to the different types of E 171, as presented in the EFSA FAF Panel (2019) opinion.
- The processes used by industry when using E 171 in food and to what extent these processes may affect the degree of agglomeration and thus internal exposure.
- State of agglomeration i.e. presence of 'free' (non-agglomerated) particles of tested material in GIT of the animals and its effect on absorption.
- Representativity of different tested materials used in toxicity and genotoxicity studies for the food additive E 171 when used in food.
- Differences in the physico-chemical properties of the different tested materials and the extent of their impact on the observed results.
- Interference in the measurements of Ti/TiO_2 in blood, tissues or organs with the most widely used analytical technique, i.e. ICP-MS, and its impact on the reliability of tissue concentration data.
- Confidence in the limited kinetic data as the basis for estimating half-lives and accumulation and for assessment of internal exposure and, related to that, the extent of systemic availability.
- None of the rodent studies were sufficiently long to cover the time needed for reaching the steady state for accumulation and this impacted the interpretation of the study results.
- Relative contribution of different molecular mechanisms leading to the production of ROS resulting in the genotoxicity of TiO₂ (inflammation, interaction with mitochondria, intrinsic potential of TiO₂ to generate ROS).
- Several modes of action for the genotoxicity may operate in parallel. The relative contributions of different molecular mechanisms elicited by TiO₂ particles are unknown; it is unclear if a threshold mode of action could be assumed.
- Nature of the interactions between DNA and TiO₂ particles leading to conformational changes in DNA



Due to large amount of information that needed reviewing in the limited time available, a structured uncertainty analysis in line with the EFSA Guidance on Uncertainty Analysis in Scientific Assessment (EFSA Scientific Committee, 2018b) was not possible. However, the Panel took a conservative approach in reaching the final conclusions.

In relation to the exposure assessment (Section 4.4.3), the Panel considered that the uncertainties identified resulted in an overestimation of the exposure to E 171 from its use as a food additive according to Annex II to Regulation No 1333/2008 for all scenarios.

6. Discussion

The safety of E 171 was re-evaluated by EFSA in 2016 in the frame of Regulation (EU) No 257/2010, as part of the re-evaluation programme for food additives authorised in the EU before 20 January 2009 (EFSA ANS Panel, 2016). On the basis of the information available at that time, the EFSA ANS Panel considered that E 171 mainly consisted of micro-sized TiO₂ particles, with a nano-sized (< 100 nm) fraction less than 3.2% by mass. Uncertainties around the identity and characterisation of E 171 were however highlighted, noting that no limits for the particle size of E 171 were set in the EU specifications. The ANS Panel concluded that, based on the data available at that time, E 171 when used as a food additive did not raise concern with respect to genotoxicity and that it was not carcinogenic after oral administration. Taking into account the presumed limited absorption of TiO₂, the ANS Panel concluded that, based on a margin of safety (MoS) calculated from the no-observed-adverse-effect level (NOAEL) of 2,250 mg TiO₂/kg bw per day (identified from a carcinogenicity study in rats) and the exposure, calculated based on the reported use levels and analytical data, E 171 would not be of concern. However, given the toxicological data set at that time, the ANS Panel identified data gaps and uncertainties that required follow-up by the European Commission by means of a call for data aimed at gathering information from IBOs. In particular, in order to address concerns related to the lack of adequate data on reproductive and developmental toxicity, the ANS Panel recommended that an EOGRT study be performed. An EOGRT study was commissioned by IBOs and its study protocol was later amended to accommodate the investigation of additional parameters related to the occurrence and TiO2-related induction of ACF in the colon, which are preneoplastic lesions that had been reported by Bettini et al. (2017) shortly after the completion of the ANS Panel re-evaluation of E 171.

Subsequent to the evaluation of data submitted by IBOs on the characterisation of E 171 used as a food additive in the EU, the Panel recommended that the EU specifications for E 171 include the parameter of median minimum external dimension by particle number should be higher than 100 nm, measured by electron microscopy, which is equivalent to less than 50% of constituent particles by number with a minimum external dimension below 100 nm (EFSA FAF Panel, 2019).

Based on the presence of a fraction of nanoparticles in E 171, the food additive falls under the scope of the EFSA Guidance on nanotechnology, which was broadened in its 2018 revision to cover also 'a material that is not engineered as nanomaterial but contains a fraction of particles, less than 50% in the number–size distribution, with one or more external dimensions in the size range 1–100 nm' (EFSA Scientific Committee, 2018a).

For the reason given above, the proposed amendment to the specifications of the food additive E 171 (EFSA FAF Panel, 2019) was accompanied by a recommendation by the Panel for a reassessment of the toxicological data set in line with the data requirements specified in the EFSA Guidance on nanotechnology (EFSA Scientific Committee, 2018a).

Scientific criteria for implementing the provisions of the EFSA Guidance on nanotechnology (EFSA Scientific Committee, 2018a) and specific information on the characteristics of TiO_2 nanoparticles were considered when preparing the advice from the ccWG Nano. The advice elaborated on the NSC and adaptations related to specific aspects of study design with TiO_2 which are adequate for a hazard identification and hazard characterisation of small particles, including nanoparticles (Appendix E). Following this advice (Appendix E), toxicokinetic and toxicity studies were scored for NSC (dispersion and/or confirmation of internal exposure). The confidence for assessing the toxicological effects of the fraction of small particles, including nanoparticles was as follows:

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- Scoring 1 for NSC: the study is suitable.
- Scoring 2 for NSC: the study has some limitations.
- Scoring 3 for NSC: the relevance of the results cannot be verified.
- Scoring 4 for NSC: the relevance of the results is low.



As mentioned above, the characterisation of E 171 was previously evaluated by the Panel who noted that E 171 currently contains nanoparticles at different percentages (EFSA FAF Panel, 2019). Moreover, from samples of E 171 or in E 171 extracted from foods the percentage by number of particles below 30 nm is in the order of 1% or less (Verleysen et al., 2020, 2021; Geiss et al., 2021, Appendix W) Therefore, the Panel considered that studies performed with TiO_2 NPs that predominantly consist of particles smaller than 30 nm (e.g. P25) are of limited relevance to the safety assessment of E 171. However, data from toxicity studies performed with $TiO_2 < 30$ nm have been considered for completeness of the database and may be relevant with respect to whether a minimum limit for particle size should be included in the EU specifications for E 171.

The Panel considered that E 171 has a low oral systemic availability, probably not higher than 0.5% (Section 4.1.2). It may pass the placenta and be transferred to the fetus. Rat studies showed long half-lives for TiO_2 NP (7–90 nm) (roughly 200–450 days), a potential for accumulation (accumulation factor of 290–450) and a long time to reach steady state (3–5 years) could be estimated from these studies. The oral systemic availability of TiO_2 NP was low (most probably < 1%) but higher than for E 171. In tissues from deceased subjects, TiO_2 particles were identified in liver, spleen, kidney and intestinal tissues. The low Ti amount of the investigated organs indicated low oral systemic availability of TiO_2 ingested from a number of sources, including dietary exposure to E 171.

The Panel noted that none of the studies were sufficiently long to cover the time needed for reaching the steady-state for accumulation. Therefore, the Panel considered that this could contribute to the uncertainty for the interpretation of the toxicological findings.

In mice, no adverse effects associated with general toxicity were observed up to 1,000 mg E 171/kg bw per day, the highest dose tested, for dosing durations up to 90 days (Han et al., 2020a, scoring 2 for NSC). Also in rats, no adverse effects associated with general toxicity were observed in the EOGRT study with E 171 (Documentation provided to EFSA No 1) (scoring 4 for NSC) at doses up to 1,000 mg/kg bw per day (Section 4.2). In rat toxicity studies with TiO_2 NPs or TiO_2 containing a fraction of nanoparticles, having different duration (14–90 days), no adverse effects were observed up to the highest dose tested (from 40 to 100 mg/kg bw per day). Overall, in a weight of evidence consideration, no adverse effects associated with general toxicity were observed in rats orally exposed to E 171, TiO_2 NPs or TiO_2 containing nanoparticles.

In mice orally exposed to TiO_2 NPs < 30 nm for up to 90 days, some effects were reported, which by their nature could be adverse. However, mild hyperbilirubinaemia was not accompanied by any in liver enzymes (Yang et al., 2017); the effect size of increased fasting glycaemia and impaired glucose tolerance (Hu et al., 2015) was not accompanied by changes in insulin or other changes in lipid metabolism and therefore was not of toxicological relevance. Histopathological changes were reported in the heart (Yu et al., 2016), however, these findings were not supported by incidences and severity scores. Histopathological findings indicating inflammation were reported in the liver but investigations to confirm hepatic injury were not performed (Hong et al., 2016).

In rats orally exposed to TiO_2 NPs < 30 nm, inconsistent and/or unexplained sex differences in some parameters were reported (e.g. hypobilirubinaemia in females (Chen et al., 2015a); heart rate and blood pressure changes in females (Chen et al., 2015b); leucocyte changes in females (Heo et al., 2020); higher absolute pituitary weights in males (Heo et al., 2020); lower blood insulin levels in females, lower C-peptide levels in males and differences in blood concentrations compared to controls in a glucose tolerance test in males (Chen et al., 2020b).

The Panel considered that the effects reported in mouse studies with TiO_2 NPs < 30 nm could be associated with accumulation of NPs in various tissues, whereas inconsistent findings in rats were considered incidental.

No effects of E 171 on sexual function and fertility in either male or female rats, and on pre- and postnatal developmental were observed in the EOGRT study with E 171 (Documentation provided o EFSA No 1, scoring 4 for NSC) up to 1,000 mg/kg bw per day, the highest dose tested (Section 4.2.7). No other reproductive or developmental toxicity studies performed with E 171 have been identified from the published literature that were considered sufficiently reliable (see Appendix H). No maternal and developmental effects were observed up to 1,000 mg/kg bw per day, the highest dose tested, in a single rat developmental toxicity study with five different TiO_2 materials, TiO_2 NPs or TiO_2 containing a fraction of nanoparticles (Warheit et al., 2015b) (scoring 4 for NSC).

In mice, the effects of TiO_2 NPs < 30 nm on the testis (decreased weight, decreased seminiferous tubule diameter, germ cell apoptosis) and sperm (decreased sperm counts and motility, increased percentage of abnormal spermatozoa) were observed in three studies (Khorsandi et al., 2016, 2017; Karimi et al., 2019) at doses ranging from 50 to 300 TiO_2 NPs mg/kg bw per day. In a mouse study by



Lu et al. (2020), no effects were observed at the lowest dose tested, 10 mg/kg bw per day (scoring 4 for NSC). In rats, administration of TiO_2 NPs (21 nm) did not show effects at any dose level in a developmental toxicity study up to 1,000 mg/kg bw per day (Lee et al., 2019) (scoring 3 for NSC).

No neurotoxicity studies performed with E 171 have been identified from the published literature that were considered sufficiently reliable (see Appendix C). Based on the results of the EOGRT study in rats with E 171 (Documentation provided to EFSA No 1, scoring 4 for NSC), the Panel considered that E 171 had no adverse effects on neurofunctional endpoints in F1 cohort 2A offspring up to 1,000 mg/kg bw per day (Section 4.2.8).

 TiO_2 NPs orally administered to rats during embryofetal and early postnatal development reduced hippocampal neurogenesis with TiO_2 NPs (< 100 nm) at 100 mg/kg bw per day exposure (Ebrahimzadeh et al., 2017), and dosed to adult rats produced adverse effects in the brain consistent with indications of oxidative stress with TiO_2 NPs (90 nm) at 500 mg/kg bw per day (Kandeil et al., 2019).

After oral dosing with TiO_2 NPs < 30 nm, adverse effects in both adult and developing mouse and rat brain were observed in studies identified from the published literature. Most of these effects are possibly related to oxidative stress. In mice, the Panel noted that the reduced volume of the polymorph layer of the hippocampal dentate gyrus and reduced density and number of dentate gyrus granular neurons reported by Rahnama et al. (2020, scoring 4 for NSC) with TiO_2 NPs (21 nm), is consistent with the behavioural effects reported by Zhang et al. (2020, scoring 3 for NSC), i.e. increased open field anxiety-like behaviour and unaffected spatial learning and memory. Ventral dentate gyrus is associated with anxiety behaviour, CA regions with spatial learning/memory (Eagle et al., 2016; Anacker et al., 2018). In adults rats, the most sensitive endpoint in the evaluated studies was reduced brain cholinesterase activity and increased brain Na/K-ATPase activity with TiO_2 NPs (21 nm) at a dose of 0.5 mg/kg bw per day in females dosed for 14 days (Canli et al., 2020, scoring 4 for NSC).

The Panel noted that inhibition of cholinesterase activity by nanoparticles other than TiO_2 , both metal and plastic, has been reported in a number of species (Prüst et al., 2020). Since oxidative stress-related inflammation is generally associated with increased and not decreased cholinesterase activity (Corrêa Mde et al., 2008; Vaknine and Soreq, 2020), it is unclear whether there is a link between TiO_2 -induced oxidative stress and TiO_2 -induced decrease in cholinesterase activity.

For neurotoxicity, adverse effects were seen with TiO_2 NPs < 30 nm. In mice, Zhou et al. (2017), reported adverse effects (i.e. inhibited dendritic outgrowth, increased autophagy and oxidative stress and reduced mitochondrial function) in *ex vivo* hippocampal neurons of weanling mice after dosing TiO_2 NPs (6–7 nm) during gestation and early lactation at a dose of 1 mg/kg bw per day, the lowest dose tested. In adult female rats (Canli et al., 2020), adverse effects (reduced brain cholinesterase activity, and increased brain Na/K-ATPase activity) were observed with TiO_2 NPs (21 nm) at 0.5 mg/kg bw per day, the lowest of three doses tested, in a 14-day study. The Panel noted that Canli et al. (2020) was scored 4 for NSC.

In the immunotoxicity cohort of the EOGRT study with E 171 (Documentation provided to EFSA No 1, scoring 4 for NSC), a slight, but statistically significant decrease (–9%) in antigen specific IgM level was measured at the highest dose tested (1,000 mg/kg bw per day) in males, but without an apparent dose response. Due to technical limitations in this part of the study, it is currently not possible to conclude on the developmental immunotoxicity of E 171 (Section 4.2.9).

From other studies identified from the published literature with E 171, the Panel concludes that these studies suggest an immune dysregulatory activity of E 171, evidenced by several immune-related and inflammatory markers.

In the Han et al. (2020a) 90-day study, effects on GM-CSF and IgM and were observed following exposure to E 171, however the Panel noted the lack of a dose response, the magnitude of the effect was small that did not allow a firm conclusion given the natural variability in the parameters measured. It should be noted that in three single dose level studies with E 171 inflammatory effects were noted at lower doses, i.e. 2, 5 and 10 mg/kg per day, respectively, in rats (Bettini et al., 2017) and in mice (Urrutia-Ortega et al., 2016; Talamini et al., 2019).

Effects of E 171 may, at least in part, stem from the activity of the fraction of the smaller TiO_2 particles, as studies with these particles also indicate inflammatory effects of exposure to TiO_2 NPs (5–6 nm) at 2.5 mg/kg per day (Yu et al., 2016).

Although not a requirement in the OECD TG 443, an evaluation of ACF in the colon of satellite F0 animals was investigated in the EOGRT study (Section 4.2.6). From this study (scoring 4 for NSC), the Panel considered that oral exposure to E 171 at doses up to 1,000 mg/kg bw per day did not induce



ACF in the colon. Two additional studies reporting information on ACF were identified from the literature search. From the study by Bettini et al. (2017) (scoring 1 for NSC), previously reviewed by the ANS Panel (EFSA ANS Panel, 2018), the Panel considered that E 171 at a dose of 10 mg/kg bw per day may induce ACF per se. In addition, E 171 enhanced ACF formation after pretreatment with a genotoxic carcinogen (i.e. DMH) in rats. From a more recent study (Blevins et al., 2019) (scoring 3 for NSC), the Panel noted that no changes in the number of ACF and ABC were observed due to E 171 exposure alone. However, limitations in the pathological examination of ABC and ACF (sampled colon area limited; technical issues with fixation) precluded a conclusion by the Panel on any potential for ABC and ACF formation. Dietary E 171, with or without treatment with DMH, had no effect on the length of the colonic glands examined or the number of goblet cells/unit.

Overall, the Panel noted that the effect of E 171 alone (without prior initiation) in producing ACF reported by Bettini et al., has not been replicated in later investigations (EOGRT and Blevins et al., 2019), but one of these investigations (Blevins et al., 2019) had methodological limitations. Furthermore, it is unclear to what extent animals were exposed to NPs in the EOGRT and Blevins et al. (2019). The Panel considered that E 171 may induce ACF in male rats at a dose of 10 mg/kg bw per day when it is dispersed in test vehicle preventing agglomeration of NPs prior to administration. The Panel noted that there is literature indicating that ACFs may be a risk factor for human colorectal cancer (Anderson et al., 2012; Drew et al., 2018; Quintanilla et al., 2019; Hong et al., 2019; Clapper et al., 2020; Kowalczyk et al., 2020; Siskova et al., 2020).

No new publications on chronic toxicity or carcinogenicity have been identified in the literature search. During the re-evaluation of E 171 in 2016, the ANS Panel had evaluated a carcinogenicity study in mice and rats (NCI, 1979), performed with TiO_2 mixed with the diet. The ANS Panel had concluded that the study indicated that TiO_2 was not carcinogenic in rats and mice. However, in the current opinion, the Panel considered that this study was not appropriate to ascertain the absence of a potential to elicit chronic toxicity and carcinogenicity by TiO_2 nanoparticles.

A number of studies, considered to be reliable by the Panel, have examined or included analyses of GIT microbiota changes in response to exposure to E 171, TiO_2 NPs and TiO_2 NPs < 30 nm. There is no consensus on quantifying the extent of GIT microbiota changes and when such changes should be considered adverse. Therefore, the Panel was unable to come to any conclusion regarding the effects of E 171 on GIT microbiota and related effects on health.

Combining the available lines of evidence, the Panel considered that TiO $_2$ particles have the potential to induce DNA strand breaks and chromosomal damage, but not gene mutations. The Panel noted that the largest particle sizes resulting in genotoxicity after oral exposure were TiO $_2$ (160 \pm 59 nm) in an *in vivo* comet assay (Sycheva et al., 2011) and TiO $_2$ NPs (58 \pm 8 nm) in *in vivo* MN and CA assays (Chakrabarti et al., 2019). Negative results were observed in two studies with particles of similar or larger size, i.e. in an *in vivo* MN assay with TiO $_2$ NPs (75 \pm 15 nm) (Chen et al., 2014) and in an *in vivo* comet assay with E 171 reported by the authors to be distributed in three size groups of particles as observed from the TEM images (135 \pm 46 nm, 305 \pm 61, 900 \pm 247 nm) but without quantitative information of each side group (Jensen et al., 2019). The Panel noted that the ranges of the particles sizes in studies resulting in a positive or negative outcome were overlapping, the number of negative studies performed with large particles is low. Accordingly, a cut-off value for TiO $_2$ particle size with respect to genotoxicity could not be identified. The Panel also considered that no clear correlation was observed between other physico-chemical properties of TiO $_2$ NPs, such as crystalline form, shape and agglomeration state and the outcome of genotoxicity assays.

The relative contribution of the MOAs, which may operate in parallel, to the resulting genotoxicity of TiO_2 is unknown and there is uncertainty on whether a thresholded mode of action could be assumed. Even if it was assumed that all potential modes of action would be indirect, the available data would not allow identification of a threshold dose. Therefore, the Panel concluded that a concern for genotoxicity of TiO_2 particles cannot be ruled out.

Considering the technical data, including heterogeneous information on particle size of E 171, as provided by IBOs during the re-evaluation of this food additive (EFSA ANS Panel, 2016), the ANS Panel considered that the highest reported percentage value of 3.2% of nanoparticles (< 100 nm) by mass, could be used as an estimate of NPs in E 171. In contrast, the current data (Verleysen et al., 2020) suggest E 171 can contain about 30% of nanoparticles < 100 nm by mass. Therefore, in the current assessment, the Panel took into consideration the principles established in the EFSA Guidance on Nanotechnology (2018). In addition, a literature review was performed for the current assessment that also captured studies performed specifically with TiO_2 NPs. Data on the potential genotoxicity of TiO_2 — that was not previously identified as relevant based on the available information on maximum



percentage value of nanoparticles present in E 171 for the 2016 re-evaluation of E 171 – were also included in the current assessment.

7. Conclusions

Concerning the content of nanoparticles in E 171, the Panel considered that:

- according to the Regulation EU (No) 231/2012, there is currently no limitation for the content of nanoparticles in E 171.
- according to data received from interested business operators, less than 50% of constituent particles in E 171 have a minimum external dimension below 100 nm by number (EFSA FAF Panel, 2019).
- the percentage by number of constituent particles below 30 nm was in the order of 1% or less in samples of pristine E 171 or in E 171 extracted from foods analysed after dispersion.
- TiO₂ particles in pristine E 171 likely form large agglomerates. When dispersion procedures are applied, these agglomerates may deagglomerate, resulting in increased numbers of 'free' nanoparticles. The extent of agglomeration and number of 'free' nanoparticles present may be further affected by the conditions in food and the GIT environment.

Accordingly, the Panel concluded that studies with TiO_2 nanoparticles were relevant in the current risk assessment of E 171. However, studies performed with TiO_2 NPs that predominantly consisted of particles smaller than 30 nm were considered to be of limited relevance.

Concerning absorption and toxicity of TiO₂ particles that are present in E 171, the Panel concluded that:

- the absorption of TiO₂ particles is low, however they can accumulate in the body due to their long half-life;
- studies on general and organ toxicity, including the newly performed EOGRT study with E 171, did not indicate adverse effects up to a dose of 1,000 mg/kg bw per day. Also, no effects were seen in studies retrieved from the literature with TiO₂ NP > 30 nm up to the highest dose tested of 100 mg/kg bw per day;
- no effects on reproductive and developmental toxicity up to a dose of 1,000 mg/kg bw per day, the highest dose tested, were observed in the EOGRT study with E 171. No other reliable studies were found in the literature addressing these effects with E 171;
- some findings regarding immunotoxicity and inflammation with E 171 as well as neurotoxicity with TiO₂ NPs may be indicative of adverse effects;
- there are indications of the induction of aberrant crypt foci with E 171;
- no studies appropriately designed and conducted to investigate the potential carcinogenicity of TiO₂ nanoparticles were available;
- combining the available lines of evidence on genotoxicity, TiO₂ particles have the potential to induce DNA strand breaks and chromosomal damage, but not gene mutations. No clear correlation was observed between the physico-chemical properties of TiO₂ particles such as crystalline form, size of constituent particles, shape and agglomeration state and the outcome of either *in vitro* or *in vivo* genotoxicity assays;
- a concern for genotoxicity of TiO₂ particles that may be present in E 171 could not be ruled out
- several modes of action for the genotoxicity may operate in parallel. The relative contributions of different molecular mechanisms elicited by TiO₂ particles are unknown and there is uncertainty whether a threshold mode of action could be assumed;
- a cut-off value for TiO₂ particle size with respect to genotoxicity could not be identified.

Overall, on the basis of all currently available evidence along with all the uncertainties, in particular the fact that genotoxicity concern could not be ruled out, the Panel concluded that E 171 can no longer be considered as safe when used as a food additive.

This conclusion applies to E 171 as described in Commission Regulation (EU) No 231/2012 as well as to E 171 specified in the EFSA FAF Panel (2019).

Documentation provided to EFSA

1) LPT (Laboratory Pharmacology and toxicology), 2020a. Interim Report - EOGRT study of Titanium dioxide E171-E in rats by oral administration via the diet. Report No. 36222.



- Submitted by the European Commission on 27 May 2020; updated version of the report submitted by Titanium Dioxide Manufacturers Association on 1st September 2020
- 2) LPT (Laboratory Pharmacology and toxicology), 2020b. Dietary Analysis EOGRT study of Titanium dioxide E 171-E in rats by oral administration via the diet. Report No. 36222, Submitted by the European Commission on 14 September 2020.
- 3) Global Pathology Support D.V, 2020. Pathology assessment EOGRT study of Titanium dioxide E 171-E in rats by oral administration via the diet (Draft amendment to final pathologist report). Test site reference no. 523/LPT Report No. 36222, Submitted by the European Commission on 15 September 2020; Draft amendment to final pathologist report submitted by Titanium Dioxide Manufacturers Association on 16 November 2020; update submitted 08 January 2021.
- 4) Fraunhofer Institute for Molecular Biology an Applied Ecology, 2020a. Determination of titanium in rat blood and urine of an EOGRT study with Titanium Dioxide E 171-E. Study number: U-EBR-244/6-27, Submitted by the European Commission on 16 September 2020.
- 5) Fraunhofer Institute for Molecular Biology an Applied Ecology, 2020b. Validation: Digestion of Titanium dioxide E 171-E in rat blood samples and determination of dissolved titanium in digested samples. Submitted by the European Commission on 16 September 2020.
- 6) Fraunhofer Institute for Molecular Biology an Applied Ecology, 2020c. Validation: Digestion of Titanium dioxide E 171-E in rat urine samples and determination of dissolved titanium in digested samples. Submitted by the European Commission on 16 September 2020.
- 7) Finish Institute of occupational health (FIOH), 2013. Facilitating the safety evaluation of manufactured nanomaterials by characterising their potential genotoxic hazard Project Coordinator French Agency for Food, Environmental and Occupational Health & Safety (ANSES) report. In vitro testing strategy for nanomaterials including database. Final report, March 2013. Submitted by FIOH, October 2020.
- 8) French Agency for Food, Environmental and Occupational Health & Safety (ANSES), 2013. Nanogenotox, Deliverable 5: In vitro testing strategy for nanomaterials including database. Final report, March 2013. Submitted by ANSES, October 2020.
- 9) National Research Centre for the Working Environment (NRCWE), 2013 NANOGENOTOX work package six report, Comet and Micronucleus in vivo data. Submitted by NRCWE, November 2020.
- 10) Federal Institute for Risk Assessment (BfR) 2012. BfR study reports of the comet for TiO_2 NP as part of the work package five of the Nanogenotox project. Submitted by BfR, November 2020.
- 11) Additional clarifications submitted in response to a request from EFSA. Submitted by Titanium Dioxide Manufacturers Association, 16th and 18th November 2020.
- 12) Additional clarifications submitted in response to a request from EFSA. Submitted by Titanium Dioxide Manufacturers Association, 7 January 2021.
- 13) Additional clarifications submitted in response to a request from EFSA. Submitted by Titanium Dioxide Manufacturers Association, 12 January 2021.
- 14) BioReliance, 2020a. Bacterial Reverse Mutation Assay. Submitted by Titanium Dioxide Manufacturers Association, 12 January 2021.
- 15) BioReliance, 2020b. In Vitro Mammalian Cell Micronucleus Assay in Human Peripheral Blood Lymphocytes (HPBL). Submitted by Titanium Dioxide Manufacturers Association, 12 January 2021.
- 16) Additional clarifications submitted in response to a request from EFSA. Submitted by Titanium Dioxide Manufacturers Association, 22 January 2021.
- 17) Additional clarifications submitted in response to a request from EFSA. Submitted by Titanium Dioxide Manufacturers Association, 29 January 2021.
- 18) Additional clarifications submitted in response to a request from EFSA. Submitted by Titanium Dioxide Manufacturers Association, 09 February 2021.

References

Ahlinder L, Ekstrand-Hammarström B, Geladi P and Osterlund L, 2013. Large uptake of titania and iron oxide nanoparticles in the nucleus of lung epithelial cells as measured by Raman imaging and multivariate classification. Biophysical Journal, 105, 310–319.

Akera T and Brody TM, 1978. The role of Na+, K+-ATPase in the inotropic action of digitalis. Pharmacological Reviews, 29, 187–220.



- Ali K, Abul QF, Dwivedi S, Abdel-Salam EM, Ansari SM, Saquib Q, Faisal M, Al-Khedhairy AA, Al-Shaeri M and Musarrat J, 2018. Titanium dioxide nanoparticles preferentially bind in subdomains IB, IIA of HSA and minor groove of DNA. Journal of Biomolecular Structure and Dynamics, 36, 2530–2542.
- Ali SA, Rizk MZ, Hamed MA, Aboul-Ela EI, El-Rigal NS, Aly HF and Abdel-Hamid AZ, 2019. Assessment of titanium dioxide nanoparticles toxicity via oral exposure in mice: effect of dose and particle size. Biomarkers: biochemical indicators of exposure, response, and susceptibility to chemicals, 24, 492–498.
- Alsudir S and Lai EPC, 2017. Electrosteric stabilization of colloidal TiO₂ nanoparticles with DNA and polyethylene glycol for selective enhancement of UV detection sensitivity in capillary electrophoresis analysis. Analytical and Bioanalytical Chemistry, 409.
- Ammendolia MG, Iosi F, Maranghi F, Tassinari R, Cubadda F, Aureli F, Raggi A, Superti F, Mantovani A and De Berardis B, 2017. Short-term oral exposure to low doses of nano-sized TiO₂ and potential modulatory effects on intestinal cells. Food and Chemical Toxicology, 102, 63–75.
- Anacker C, Luna VM, Stevens GS, Millette A, Shores R, Jimenez JC, Chen B and Hen R, 2018. Hippocampal neurogenesis confers stress resilience by inhibiting the ventral dentate gyrus. Nature, 559, 98–102.
- Anderson JC, Swede H, Rustagi T, Protiva P, Pleau D, Brenner BM, Rajan TV, Heinen CD, Levine JB and Rosenberg DW, 2012. Aberrant crypt foci as predictors of colorectal neoplasia on repeat colonoscopy. Cancer Causes & Control, 23, 355–361.
- Andreoli C, Leter G, De Berardis B, Degan P, De Angelis I, Pacchierotti F, Crebelli R, Barone F and Zijno A, 2018. Critical issues in genotoxicity assessment of TiO₂ nanoparticles by human peripheral blood mononuclear cells. Journal of Applied Toxicology: JAT, 38, 1471–1482.
- ANSES (Agence nationale de sécurité sanitaire de l'alimentation, de l'environnement et du travail), 2017. Relatif à une demande d'avis relatif à l'exposition alimentaire aux nanoparticules de dioxyde de titane. Referral No. 2017-SA-0020.
- ANSES (Agence nationale de sécurité sanitaire de l'alimentation, de l'environnement et du travail), 2019. Relatif aux risques liés à l'ingestion de l'additif alimentaire E 171. Referral No. 2019-SA-0036.
- Asare N, Duale N, Slagsvold HH, Lindeman B, Olsen AK, Gromadzka-Ostrowska J, Meczynska-Wielgosz S, Kruszewski M, Brunborg G and Instanes C, 2016. Genotoxicity and gene expression modulation of silver and titanium dioxide nanoparticles in mice. Nanotoxicology, 10, 312–321.
- Azim SA, Darwish HA, Rizk MZ, Ali SA and Kadry MO, 2015. Amelioration of titanium dioxide nanoparticles-induced liver injury in mice: possible role of some antioxidants. Experimental and Toxicologic Pathology: Official Journal of the Gesellschaft fur Toxikologische Pathologie, 67, 305–314.
- Bajic V, Spremo-Potparevic B, Zivkovic L, Cabarkapa A, Kotur-Stevuljevic J, Isenovic E, Sredojevic D, Vukoje I, Lazic V, Ahrenkiel SP and Nedeljkovic JM, 2017. Surface-modified TiO₂ nanoparticles with ascorbic acid: antioxidant properties and efficiency against DNA damage in vitro. Colloids and surfaces B, Biointerfaces, 155, 323–331.
- Barillet S, Simon-Deckers A, Herlin-Boime N, Mayne-L'Hermite M, Reynaud C, Cassio D, Gouget B and Carriere M, 2010. Toxicological consequences of TiO₂, SiC nanoparticles and multi-walled carbon nanotubes exposure in several mammalian cell types: an in vitro study. Journal of Nanoparticle Research, 12, 61–73.
- Barkhade T, Mahapatra KS and Banerjee I, 2019. Study of mitochondrial swelling, membrane fluidity and ROS production induced by nano-TiO₂ and prevented by Fe incorporation. Toxicol. Res, 8, 711.
- Bayat N, Lopes VR, Scholermann J, Jensen LD and Cristobal S, 2015. Vascular toxicity of ultra-small TiO₂ nanoparticles and single walled carbon nanotubes in vitro and in vivo. Biomaterials, 63, 1–13.
- Bedard K and Krause K-H, 2007. The NOX family of ROS-Generating NADPH oxidases: physiology and pathophysiology. Physiological Reviews, 87, 245–313.
- Bettini S, Boutet-Robinet E, Cartier C, Comera C, Gaultier E, Dupuy J, Naud N, Tache S, Grysan P, Reguer S, Thieriet N, Refregiers M, Thiaudiere D, Cravedi JP, Carriere M, Audinot JN, Pierre FH, Guzylack-Piriou L and Houdeau E, 2017. Food-grade TiO₂ impairs intestinal and systemic immune homeostasis, initiates preneoplastic lesions and promotes aberrant crypt development in the rat colon. Scientific Reports, 7, 40373.
- Bhattacharya K, Davoren M, Boertz J, Schins RPF, Hoffmann E and Dopp E, 2009. Titanium dioxide nanoparticles induce oxidative stress and DNA-adduct formation but not DNA-breakage in human lung cells. Particle and Fibre Toxicology, 6, 17.
- Biola-Clier M, Beal D, Caillat S, Libert S, Armand L, Herlin-Boime N, Sauvaigo S, Douki T and Carriere M, 2017. Comparison of the DNA damage response in BEAS-2B and A549 cells exposed to titanium dioxide nanoparticles. Mutagenesis, 32, 161–172.
- Blevins LK, Crawford RB, Bach A, Rizzo MD, Zhou J, Henriquez JE, Khan D, Sermet S, Arnold LL, Pennington KL, Souza NP, Cohen SM and Kaminski NE, 2019. Evaluation of immunologic and intestinal effects in rats administered an E 171-containing diet, a food grade titanium dioxide (TiO₂). Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association, 133.
- Brandão F, Fernández-Bertólez N, Rosário F, Bessa MJ, Fraga S, Pásaro E, Teixeira JP, Laffon B, Valdiglesias V and Costa C, 2020. Genotoxicity of TiO₂ nanoparticles in four different human cell lines (A549, HEPG2, A172 and SH-SY5Y). Nanomaterials, Basel, 10.



- Brown DM, Danielsen PH, Derr R, Moelijker N, Fowler P, Stone V, Hendriks G, Moller P and Kermanizadeh A, 2019. The mechanism-based toxicity screening of particles with use in the food and nutrition sector via the ToxTracker reporter system. Toxicology in vitro: an international journal published in association with BIBRA, 61.
- Brzicova T, Javorkova E, Vrbova K, Zajicova A, Holan V, Pinkas D, Philimonenko V, Sikorova J, Klema J, Topinka J and Rossner Jr P, 2019. Molecular responses in THP-1 macrophage-like cells exposed to diverse nanoparticles. Nanomaterials (Basel, Switzerland). 9 pp.
- Cani PD and Everard A, 2015. Keeping gut lining at bay: impact of emulsifiers. Trends in Endocrinology and Metabolism, 26, 273–274.
- Canli EG, Gumus C, Canli M and Ila HB, 2020. The effects of titanium nanoparticles on enzymatic and non-enzymatic biomarkers in female Wistar rats. Drug and Chemical Toxicology, 1–9.
- Chakrabarti S, Goyary D, Karmakar S and Chattopadhyay P, 2019. Exploration of cytotoxic and genotoxic endpoints following sub-chronic oral exposure to titanium dioxide nanoparticles. Toxicology and Industrial Health, 35, 577–592.
- Charles S, Jomini S, Fessard V, Bigorgne-Vizade E, Christophe Rousselle C and Michel C, 2018. Assessment of the *in vitro* genotoxicity of TiO₂ nanoparticles in a regulatory context. Nanotoxicology, 12, 357–374. https://doi.org/10.1080/17435390.2018.1451567
- Chassaing B, Koren O, Goodrich JK, Poole AC, Srinivasan S, Ley RE and Gewirtz AT, 2015. Dietary emulsifiers impact the mouse gut microbiota promoting colitis and metabolic syndrome. Nature, 519, 92–96.
- Chassaing B, de Wiele TV, De Bodt J, Marzorati M and Gewirtz X, 2017. Dietary emulsifiers directly alter human microbiota composition and gene expression ex vivo potentiating intestinal inflammation. Gut, 1–14.
- Chen J, Dong X, Zhao J and Tang G, 2009. In vivo acute toxicity of titanium dioxide nanoparticles to mice after intraperitoneal injection. Journal of Applied Toxicology, 29, 330.
- Chen X-X, Cheng B, Yang Y-X, Cao A, Liu J-H, Du L-J, Liu Y, Zhao Y and Wang H, 2013. Characterization and preliminary toxicity assay of nano-titanium dioxide additive in sugar-coated chewing gum. Small (Weinheim an der Bergstrasse, Germany), 9, 1765–1774. https://doi.org/10.1002/smll.201201506
- Chen Z, Wang Y, Ba T, Li Y, Pu J, Chen T, Song Y, Gu Y, Qian Q, Yang J and Jia G, 2014. Genotoxic evaluation of titanium dioxide nanoparticles in vivo and in vitro. Toxicology Letters, 226, 314–319.
- Chen Z, Wang Y, Zhuo L, Chen S, Zhao L, Chen T, Li Y, Zhang W, Gao X, Li P, Wang H and Jia G, 2015a. Interaction of titanium dioxide nanoparticles with glucose on young rats after oral administration. Nanomedicine: Nanotechnology, Biology, and Medicine, 11, 1633–1642.
- Chen Z, Wang Y, Zhuo L, Chen S, Zhao L, Luan X, Wang H and Jia G, 2015b. Effect of titanium dioxide nanoparticles on the cardiovascular system after oral administration. Toxicology Letters, 239, 123–130.
- Chen Z, Han S, Zhou D, Zhou S and Jia G, 2019. Effects of oral exposure to titanium dioxide nanoparticles on gut microbiota and gut-associated metabolism in vivo. Nanoscale, 11, 22398–22412.
- Chen Z, Zheng P, Han S, Zhang J, Li Z, Zhou S and Jia G, 2020a. Tissue-specific oxidative stress and element distribution after oral exposure to titanium dioxide nanoparticles in rats. Nanoscale, 12, 20033–20046.
- Chen Z, Han S, Zheng P, Zhou D, Zhou S and Jia G, 2020b. Effect of oral exposure to titanium dioxide nanoparticles on lipid metabolism in Sprague-Dawley rats. Nanoscale, 12, 5973–5986.
- Clapper ML, Chang WL and Cooper HS, 2020. Dysplastic aberrant crypt foci: biomarkers of early colorectal neoplasia and response to preventive intervention. Cancer Prevention Research (Philadelphia, Pa.), 13, 229–240.
- Coméra C, Cartier C, Gaultier E, Catrice O, Panouille Q, El Hamdi S, Tirez K, Nelissen I, Théodorou V and Houdeau E, 2020. Jejunal villus absorption and paracellular tight junction permeability are major routes for early intestinal uptake of food-grade TiO₂ particles: an in vivo and ex vivo study in mice. Part Fibre Toxicol, 17, 26.
- Corrêa Mde C, Maldonado P, da Rosa CS, Lunkes G, Lunkes DS, Kaizer RR, Ahmed M, Morsch VM, Pereira ME and Schetinger MR, 2008. Oxidative stress and erythrocyte acetylcholinesterase (AChE) in hypertensive and ischemic patients of both acute and chronic stages. Biomedicine & Pharmacotherapy, 62, 317–324.
- Cowie H, Magdolenova Z, Saunders M, Drlickova M, Correia CS, Halamoda KB, Gombau L, Guadagnini R, Lorenzo Y, Walker L, Fjellsbo LM, Huk A, Rinna A, Tran L, Volkovova K, Boland S, Juillerat-Jeanneret L, Marano F, Collins AR and Dusinska M, 2015. Suitability of human and mammalian cells of different origin for the assessment of genotoxicity of metal and polymeric engineered nanoparticles. Nanotoxicology, 9, 57–65.
- Cupi D and Baun A, 2016. Methodological considerations for using umu assay to assess photo-genotoxicity of engineered nanoparticles. Mutation Research Genetic Toxicology and Environmental Mutagenesis, 796, 34–39.
- Danielsen PH, Knudsen KB, Strancar J, Umek P, Koklic T, Garvas M, Vanhala E, Savukoski S, Ding Y, Madsen AM, Jacobsen NR, Weydahl IK, Berthing T, Poulsen SS, Schmid O, Wolff H and Vogel U, 2020. Effects of physicochemical properties of TiO₂ nanomaterials for pulmonary inflammation, acute phase response and alveolar proteinosis in intratracheally exposed mice. Toxicology and Applied Pharmacology, 386.
- Dankovic D, Kuempel E and Wheeler M, 2007. An approach to risk assessment for TiO₂. InhALTion Toxicology, 19, 205–212.
- Dekanski D, Spremo-Potparevic B, Bajic V, Zivkovic L, Topalovic D, Sredojevic DN, Lazic V and Nedeljkovic JM, 2018. Acute toxicity study in mice of orally administrated TiO_2 nanoparticles functionalized with caffeic acid. Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association, 115, 42–48.



- Demir E, Burgucu D, Turna F, Aksakal S and Kaya B, 2013. Determination of TiO₂, ZrO₂, and Al₂O₃ nanoparticles on genotoxic responses in human peripheral blood lymphocytes and cultured embyronic kidney cells. Journal of Toxicology and Environmental Health, Part A: Current Issues, 76, 990–1002.
- Demir E, Akca H, Turna F, Aksakal S, Burgucu D, Kaya B, Tokgun O, Vales G, Creus A and Marcos R, 2015. Genotoxic and cell-transforming effects of titanium dioxide nanoparticles. Environmental Research, 136, 300–308.
- Demir E, Creus A and Marcos R, 2017. Titanium dioxide and zinc oxide nanoparticles are not mutagenic in the mouse. Fresenius Environmental Bulletin, 26, 1001–1016.
- Di Bucchianico S, Cappellini F, Le Bihanic F, Zhang Y, Dreij K and Karlsson HL, 2017. Genotoxicity of TiO₂ nanoparticles assessed by mini-gel comet assay and micronucleus scoring with flow cytometry. Mutagenesis, 32, 127–137.
- Di Virgilio AL, Reigosa M, Arnal PM, Fernandez L and de Mele M, 2010. Comparative study of the cytotoxic and genotoxic effects of titanium oxide and aluminium oxide nanoparticles in Chinese hamster ovary (CHO-K1) cells. Journal of Hazardous Materials, 177, 711–718.
- Disdier C, Devoy J, Cosnefroy A, Chalansonnet M, Herlin-Boime N, Brun E, Lund A and Mabondzo A, 2015. Tissue biodistribution of intravenously administrated titanium dioxide nanoparticles revealed blood-brain barrier clearance and brain inflammation in rat. Particle and Fibre Toxicology, 12, 24.
- Dobrzynska MM, Gajowik A, Radzikowska J, Lankoff A, Dusinska M and Kruszewski M, 2014. Genotoxicity of silver and titanium dioxide nanoparticles in bone marrow cells of rats in vivo. Toxicology, 315, 86–91.
- Donner EM, Myhre A, Brown SC, Boatman R and Warheit DB, 2016. In vivo micronucleus studies with 6 titanium dioxide materials (3 pigment-grade & 3 nanoscale) in orally-exposed rats. Regulatory Toxicology and Pharmacology: RTP, 74, 64–74.
- Dorier M, Brun E, Veronesi G, Barreau F, Pernet-Gallay K, Desvergne C, Rabilloud T, Carapito C, Herlin-Boime N and Carriere M, 2015. Impact of anatase and rutile titanium dioxide nanoparticles on uptake carriers and efflux pumps in Caco-2 gut epithelial cells. Nanoscale, 7, 7352–7360.
- Dorier M, Beal D, Marie-Desvergne C, Dubosson M, Barreau F, Houdeau E, Herlin-Boime N and Carriere M, 2017. Continuous in vitro exposure of intestinal epithelial cells to E171 food additive causes oxidative stress, inducing oxidation of DNA bases but no endoplasmic reticulum stress. Nanotoxicology, 11, 751–761.
- Dorier M, Tisseyre C, Dussert F, Beal D, Arnal ME, Douki T, Valdiglesias V, Laffon B, Fraga S, Brandao F, Herlin-Boime N, Barreau Rabilloud T and Carriere M, 2019. Toxicological impact of acute exposure to E171 food additive and TiO₂ nanoparticles on a co-culture of Caco-2 and HT29-MTX intestinal cells. Mutation Research, 845.
- Dostanic I, Schultz Jel J, Lorenz JN and Lingrel JB, 2004. The alpha 1 isoform of Na, K-ATPase regulates cardiac contractility and functionally interacts and co-localizes with the Na/Ca exchanger in heart. Journal of Biological Chemistry, 279, 54053–54061.
- Drew DA, Mo A, Grady JJ, Stevens RG, Levine JB, Brenner BM, Anderson JC, Forouhar F, O'Brien MJ, Devers TJ and Rosenberg DW, 2018. Proximal aberrant crypt foci associate with synchronous neoplasia and are primed for neoplastic progression. Molecular Cancer Research, 16, 486–495.
- Driscoll KE, Deyo LC, Carter JM, Howard BW, Hassenbein DG and Bertram TA, 1997. Effects of particle exposure and particle-elicited inflammatory cells on mutation in rat alveolar epithelial cells. Carcinogenesis, 18, 423–430.
- Du X, Gao S, Hong L, Zheng X, Zhou Q and Wu J, 2019. Genotoxicity evaluation of titanium dioxide nanoparticles using the mouse lymphoma assay and the Ames test. Mutation Research Genetic Toxicology and Environmental Mutagenesis, 838, 22–27.
- Dunkel VC, Zeiger E, Brusick D, McCoy E, McGregor D, Mortelmans K, Rosenkranz HS and Simmon VF, 1985. Reproducibility of microbial mutagenicity assays: II. Testing of carcinogens and noncarcinogens in *Salmonella typhimurium* and *Escherichia coli*. Environmental MutageneisMutagenesis, 7(Suppl 5), 1–248.
- Eagle AL, Wang H and Robison AJ, 2016. Sensitive assessment of hippocampal learning using temporally dissociated passive avoidance task. Bio Protoc, 6.
- Ebrahimzadeh BA, Mohammadipour A, Fazel A, Haghir H, Rafatpanah H, Hosseini M and Rajabzadeh A, 2017. Maternal exposure to titanium dioxide nanoparticles during pregnancy and lactation alters offspring hippocampal mRNA BAX and Bcl-2 levels, induces apoptosis and decreases neurogenesis. Experimental and toxicologic pathology: official journal of the Gesellschaft fur Toxikologische Pathologie, 69, 329–337.
- ECHA (European Chemical Agency), 2011. Guidance on information requirements and chemical safety assessment. Chapter R.4: Evaluation of available information. https://echa.europa.eu/documents/10162/13643/information_requirements_r4_en.pdf/d6395ad2-1596-4708-ba86-0136686d205e
- ECHA (European Chemicals Agency), 2017. Committee for risk assessment RAC opinion proposing harmonised classification and labelling at EU level of titanium dioxide. EC Number: 236-675-5.
- EFSA (European Food Safety Authority), 2007. Opinion of the Scientific Committee related to Uncertainties in Dietary Exposure Assessment. EFSA Journal 2007;5(1):438, 54 pp. https://doi.org/10.2903/j.efsa.2007.438
- EFSA (European Food Safety Authority), 2011. Use of the EFSA Comprehensive European Food Consumption Database in Exposure Assessment. EFSA Journal 2011;9(3):2097, 34 pp. https://doi.org/10.2903/j.efsa.2011.2097
- EFSA (European Food Safety Authority), 2015. The food classification and description system FoodEx2 (revision 2). EFSA Supporting Publication 2015;12(5):EN-804, 90 pp. https://doi.org/10.2903/sp.efsa.2015.EN-804



- EFSA (European Food Safety Authority), 2019. statement on the review of the risks related to the exposure to the food additive titanium dioxide (E 171) performed by the French Agency for Food, Environmental and Occupational Health and Safety (ANSES). EFSA Journal 2019;17(6):5714, 11 pp. https://doi.org/10.2903/j.efsa. 2019.5714
- EFSA ANS Panel (EFSA Panel on Food Additives and Nutrients Sources added to Food), 2016. Re-evaluation of titanium dioxide (E 171) as a food additive. EFSA Journal 2016;14(9):4545, 83 pp. https://doi.org/10.2903/j.efsa.2016.4545
- EFSA ANS Panel (EFSA Panel on Food Additives and Nutrients Sources added to Food), 2017. Approach followed for the refined exposure assessment as part of the safety assessment of food additives under re-evaluation. EFSA Journal 2017;15(10):5042, 9 pp. https://doi.org/10.2903/j.efsa.2017.5042
- EFSA ANS Panel (EFSA Panel on Food Additives and Nutrients Sources added to Food), 2018. Evaluation of four new studies on the potential toxicity of titanium dioxide used as a food additive (E 171). EFSA Journal 2018;16 (7):5366, 27 pp. https://doi.org/10.2903/j.efsa.2018.5366
- EFSA FAF Panel (EFSA Panel on Food Additive and Flavourings), 2019. Scientific opinion on the proposed amendment of the EU specification for titanium dioxide (E 171) with respect to the inclusion of additional parameters related to its particle size distribution. EFSA Journal 2019;17(7):5760, 23 pp. https://doi.org/10.2903/j.efsa.2019.5760
- EFSA Scientific Committee, 2009. Guidance of the scientific Committee on transparency in the scientific aspects of risk assessments carried out by EFSA. Part 2: general principles. EFSA Journal 2009;7(7):1051, 22 pp. https://doi.org/10.2903/j.efsa.2009.1051
- EFSA Scientific Committee, 2011a. Scientific Opinion on Guidance on the risk assessment of the application of nanoscience and nanotechnologies in the food and feed chain. EFSA Journal 2011;9(5):2140, 36 pp. https://doi.org/10.2903/j.efsa.2011.2140
- EFSA Scientific Committee, 2011b. Scientific Opinion on genotoxicity testing strategies applicable to food and feed safety assessment. EFSA Journal 2011;9(9):2379, 69 pp. https://doi.org/10.2903/j.efsa.2011.2379
- EFSA Scientific Committee, 2017. Scientific Opinion on the clarification of some aspects related to genotoxicity assessment. EFSA Journal 2017;15(12):5113, 25 pp. https://doi.org/10.2903/j.efsa.2017.5113
- EFSA Scientific Committee, 2018a. Guidance on risk assessment of the application of nanoscience and nanotechnologies in the food and feed chain: part 1, human and animal health. EFSA Journal 2018;16(7):5327, 95 pp. https://doi.org/10.2903/j.efsa.2018.5327
- EFSA Scientific Committee, 2018b. Guidance on uncertainty analysis in scientific assessments. EFSA Journal 2018;16(1):5123, 35 pp. https://doi.org/10.2903/j.efsa.2018.5123
- EFSA Scientific Committee, 2020. Public consultation on the draft EFSA Guidance on technical requirements for regulated food and feed product applications to establish the presence of small particles including nanoparticles. Available online: https://www.efsa.europa.eu/en/consultations/call/public-consultation-draft-efsaguidance-technical-requirements
- El Yamani N, Collins AR, Runden-Pran E, Fjellsbo LM, Shaposhnikov S, Zienolddiny S and Dusinska M, 2017. In vitro genotoxicity testing of four reference metal nanomaterials, titanium dioxide, zinc oxide, cerium oxide and silver: towards reliable hazard assessment. Mutagenesis, 32, 117–126.
- El-Bassyouni GT, Eshak MG, Barakat IAH and Khalil WKB, 2017. Immunotoxicity evaluation of novel bioactive composites in male mice as promising orthopaedic implants. Central-European Journal of Immunology, 42, 54–67.
- El-Din EAA, Mostafa HE, Samak MA, Mohamed EM and El-Shafei DA, 2019. Could curcumin ameliorate titanium dioxide nanoparticles effect on the heart? A histopathological, immunohistochemical, and genotoxic study. Environmental Science and Pollution Research International, 26, 21556–21564.
- Elespuru R, Pfuhler S, Aardema MJ, Chen T, Doak SH, Doherty A, Farabaugh CS, Kenny J, Manjanatha M, Mahadevan B, Moore MM, Ouédraogo G, Stankowski LF Jr and Tanir JY, 2018. Genotoxicity assessment of nanomaterials: recommendations on best practices, assays, and methods. Toxicological Sciences, 164, 391–416. https://doi.org/10.1093/toxsci/kfy100
- El-Ghor AA, Noshy MM, Gala A and Mohamed HRH, 2014. Normalization of nano-sized TiO₂-induced clastogenicity, genotoxicity and mutagenicity by chlorophyllin administration in mice brain, liver, and bone marrow cells. Toxicological Sciences, 142, 21–32.
- Elje E, Mariussen E, Moriones OH, Bastús NG, Puntes V, Kohl Y, Dusinska M and Rundén-Pran E, 2020. Hepato (geno)toxicity assessment of nanoparticles in a HEPG2 liver spheroid model. Nanomaterials, Basel, 10.
- Elnagar AMB, Ibrahim A and Soliman AM, 2018. Histopathological effects of titanium dioxide nanoparticles and the possible protective role of N-acetylcysteine on the testes of male albino rats. International Journal of Fertility & Sterility, 12, 249–256.
- Emi T, Rivera LM, Tripathi VC, Yano N, Ragavendran A, Wallace J and Fedulov A, 2020. Transcriptomic and epigenomic effects of insoluble particles on J774 macrophages. Epigenetics, 1–18.
- Fadda LM, Hagar H, Mohamed AM and Ali HM, 2018. Quercetin and idebenone ameliorate oxidative stress, inflammation, DNA damage, and apoptosis induced by titanium dioxide nanoparticles in rat liver. Dose-Response: A Publication of International Hormesis Society, 16, 1559325818812188.



- Fadda LM, Ali HM, Mohamed AM and Hagar H, 2019. Prophylactic administration of carnosine and melatonin abates the incidence of apoptosis, inflammation, and DNA damage induced by titanium dioxide nanoparticles in rat livers. Environmental Science and Pollution Research International.
- Fadoju O, Ogunsuyi O, Akanni O, Alabi O, Alimba C, Adaramoye O, Cambier S, Eswara S, Gutleb AC and Bakare A, 2019. Evaluation of cytogenotoxicity and oxidative stress parameters in male Swiss mice co-exposed to titanium dioxide and zinc oxide nanoparticles. Environmental Toxicology and Pharmacology, 70.
- Falck GCM, Lindberg HK, Suhonen S, Vippola M, Vanhala E, Catalan J, Savolainen K and Norppa H, 2009. Genotoxic effects of nanosized and fine TiO₂. Human and Experimental Toxicology, 28, 339–352.
- Feng L, Zhang Y, Jiang M, Mo Y, Wan R, Jia Z, Tollerud DJ, Zhang X and Zhang Q, 2015. Up-regulation of Gadd45αalpha after exposure to metal nanoparticles: the role of hypoxia inducible factor 1αalpha. Environmental Toxicology, 30, 490–499.
- Fenoglio I, Greco G, Livraghi S and Fubini B, 2009. Non-UV-induced radical reactions at the surface of TiO₂ nanoparticles that may trigger toxic responses. Chemistry—A European Journal, 15, 4614–4621.
- Ferraro D, Anselmi-Tamburini U, Tredici IG, Ricci V and Sommi P, 2016. Overestimation of nanoparticles-induced DNA damage determined by the comet assay. Nanotoxicology, 10, 861–870.
- Figtree GA, Keyvan Karimi G, Liu CC and Rasmussen HH, 2012. Oxidative regulation of the Na(+)-K(+) pump in the cardiovascular system. Free Radical Biology and Medicine, 53, 2263–2268.
- Franchi LP, Manshian BB, de Souza TA, Soenen SJ, Matsubara EY, Rosolen JM and Takahashi CS, 2015. Cyto- and genotoxic effects of metallic nanoparticles in untransformed human fibroblast. Toxicology in Vitro: An International Journal Published in ASSOCIATION with BIBRA, 29, 1319–1331.
- Franz P, Bürkle A, Wick P and Hirsch C, 2020. Exploring flow cytometry-based micronucleus scoring for reliable nanomaterial genotoxicity assessment. Chemical Research in Toxicology.
- Gallagher J, Heinrich U, George M, Hendee L, Phillips DH and Lewtas J, 1994. Formation of DNA adducts in rat lung following chronic inhALTion of diesel emissions, carbon black and titanium dioxide particles. Carcinogenesis, 15, 1291–1299.
- Garcia-Rodriguez A, Vila L, Cortes C, Hernandez A and Marcos R, 2018. Effects of differently shaped TiO₂NPs (nanospheres, nanorods and nanowires) on the in vitro model (Caco-2/HT29) of the intestinal barrier. Particle and Fibre Toxicology, 15, 33.
- Geiss O, Bianchi I, Senaldi C, Bucher G, Verleysen E, Waegeneers N, Brassinne F, Mast J, Loeschner K, Vidmar J, Aureli F, Cubadda F, Raggi A, Iacoponi F, Peters R, Undas A, Müller A, Meinhardt AK, Walz E, Gräf V and Barrero-Moreno J, 2021. Particle size analysis of pristine food-grade titanium dioxide and E 171 in confectionery products: Interlaboratory testing of a single-particle inductively coupled plasma mass spectrometry screening method and confirmation with transmission electron microscopy. Food Control, 120, 107550. https://doi.org/10.1016/j.foodcont.2020.107550. PMID: 33536722; PMCID: PMC7730118
- Geraets L, Oomen AG, Krystek P, Jacobsen NR, Wallin H, Laurentie M, Verharen HW, Brandon EF and de Jong WH, 2014. Tissue distribution and elimination after oral and intravenous administration of different titanium dioxide nanoparticles in rats. Part Fibre Toxicol, 11, 30.
- Ghosh M, Chakrabortyb A and Mukherjeea A, 2013. Cytotoxic, genotoxic and the hemolytic effect of titanium dioxide (TiO₂) nanoparticles on human erythrocyte and lymphocyte cells in vitro. Journal of Applied Toxicology.
- Gilmour P, Brown DM, Beswick PH, Benton E, MacNee W and Donaldson K, 1997. Surface free radical activity of PM10 and ultrafine titanium dioxide: a unifying factor in their toxicity? The Annals of Occupational Hygiene, 41, 32–38.
- Gore ER, Gower J, Kurali E, Sui JL, Bynum J, Ennulat D and Herzyk DJ, 2004. Primary antibody response to keyhole limpet hemocyanin in rat as a model for immunotoxicity evaluation. Toxicology, 197, 23–35.
- Griffiths PJ, Isackson H, Pelc R, Redwood CS, Funari SS, Watkins H and Ashley CC, 2009. Synchronous in situ ATPase activity, mechanics, and Ca2+ sensitivity of human and porcine myocardium. Biophysical Journal, 97, 2503–2512.
- Grissa I, Elghoul J, Ezzi L, Chakroun S, Kerkeni E, Hassine M, El Mir L, Mehdi M, Ben Cheikh H and Haouas Z, 2015. Anemia and genotoxicity induced by sub-chronic intragastric treatment of rats with titanium dioxide nanoparticles. Mutation Research Genetic Toxicology and Environmental Mutagenesis, 794, 25–31.
- Grissa I, Guezguez S, Ezzi L, Chakroun S, Sallem A, Kerkeni E, Elghoul J, El Mir L, Mehdi M, Cheikh HB and Haouas Z, 2016. The effect of titanium dioxide nanoparticles on neuroinflammation response in rat brain. Environmental Science and Pollution Research International, 23, 20205–20213.
- Grissa I, Ezzi L, Chakroun S, Mabrouk A, Saleh AB, Braham H, Haouas Z and Cheikh HB, 2017. *Rosmarinus officinalis* L. ameliorates titanium dioxide nanoparticles and induced some toxic effects in rats' blood. Environmental Science and Pollution Research International, 24, 12474–12483.
- Grissa I, ElGhoul J, Mrimi R, Mir LE, Cheikh HB and Horcajada P, 2020. In deep evaluation of the neurotoxicity of orally administered TiO₂ nanoparticles. Brain Research Bulletin, 155, 119–128.
- Guichard Y, Schmit J, Darne C, Gate L, Le Goutet M, Rousset D, Rastoix O wrobel R, Witschger O, Martin A, Fierro V and Binet S, 2012. Cytotoxicity and genotoxicity of nanosized and microsized titanium dioxide and iron oxide particles in Syrian hamster embryo cells. Annals of Occupational Hygiene, 56, 631–644.



- Guillard A, Gaultier E, Cartier C, Devoille L, Noireaux J, Chevalier L, Morin M, Grandin F, Lacroix MZ, Coméra C, Cazanave A, de Place A, Gayrard V, Bach V, Chardon K, Bekhti N, Adel-Patient K, Vayssière C, Fisicaro P, Feltin N, de la Farge F, Picard-Hagen N, Lamas B and Houdeau E, 2020. Basal Ti level in the human placenta and meconium and evidence of a materno-foetal transfer of food-grade TiO₂ nanoparticles in an ex vivo placental perfusion model. Part Fibre Toxicol, 17, 51.
- Guiot C and Spalla O, 2013. Stabilization of TiO₂ nanoparticles in complex medium through a pH adjustment protocol. Environmental Science and Technology, 15, 47.
- Gurr JR, Wang AS, Chen CH and Jan KY, 2005. Ultrafine titanium dioxide particles in the absence of photoactivation can induce oxidative damage to human bronchial epithelial cells. Toxicology, 15, 66–73.
- Hackenberg S, Friehs G, Kessler M, Froelich K, Ginzkey C, Koehler C, Scherzed A, Burghartz M and Kleinsasser N, 2011. Nanosized titanium dioxide particles do not induce DNA damage in human peripheral blood lymphocytes. Environmental and Molecular Mutagenesis, 52, 264–268.
- Haleem AM, Abbas RH, Jawad MA and Alberaqdar F, 2019. Cytotoxic effects of titanium dioxide nanaoparticles synthesized by laser technique on peripheral blood lymphocytes and hep-2 Cell Line. Toxicology and Environmental Health Sciences, 11, 219–225.
- Han HY, Yang MJ, Yoon C, Lee GH, Kim DW, Kim TW, Kwak M, Heo MB, Lee TG, Kim S, Oh JH, Lim HJ, Oh I, Yoon S and Park EJ, 2020a. Toxicity of orally administered food-grade titanium dioxide nanoparticles. Journal of Applied Toxicology.
- Han B, Pei Z, Shi L, Wang Q, Li C, Zhang B, Su X, Zhang N, Zhou L, Zhao B, Niu Y and Zhang R, 2020b. TiO₂ nanoparticles caused DNA damage in lung and extra-pulmonary organs through ROS- activated FOXO3a signaling pathway after intratracheal administration in rats. Int J Nanomedicine, 15, 6279–6294.
- Hashem MM, Abo-El-Sooud K, Abd-Elhakim YM, Badr YA, El-Metwally AE and Bahy-El-Dien A, 2020. The long-term oral exposure to titanium dioxide impaired immune functions and triggered cytotoxic and genotoxic impacts in rats. Journal of Trace Elements in Medicine and Biology, 60.
- Hassanein KM and El-Amir YO, 2017. Protective effects of thymoquinone and avenanthramides on titanium dioxide nanoparticles induced toxicity in Sprague-Dawley rats. Pathology, Research and Practice, 213, 13–22.
- Hekmat A, Saboury AA, Divsalar A and Seyedarabi A, 2013. Structural effects of TiO₂ nanoparticles and doxorubicin on DNA and their antiproliferative roles in T47D and MCF7 cells. Anti-Cancer Agents in Medicinal Chemistry, 13.
- Hekmat A, Afrough M, Tackallou SH and Ahmad F, 2020. Synergistic effects of Titanium dioxide nanoparticles and Paclitaxel combination on the DNA structure and their antiproliferative role on MDA-MB-231 cells. Journal of Nanoanalysis, 7, 152–165.
- Hendrickson OD, Pridvorova SM, Zherdev AV, Klochkov SG, Novikova OV, Shevtsova EF, Bachurin SO and Dzantiev BB, 2016. Size-dependent differences in biodistribution of titanium dioxide nanoparticles after sub-acute intragastric administrations to rats. Current Nanoscience, 12, 228–236.
- Hendrickson OD, Platonova TA, Piidvorova SM, Zherdev AV, Gmoshinsky IV, Vasilevskaya LS, Shumakova AA, Hotimchenko SA and Dzantiev BB, 2020. Electron-microscopic investigation of the distribution of titanium dioxide (rutile) nanoparticles in the rats' small intestine mucosa. Liver, and Spleen Current Nanoscience, 16, 268–279.
- Heo MB, Kwak M, An KS, Kim HJ, Ryu HY, Lee SM, Song KS, Kim IY, Kwon JH and Lee TG, 2020. Oral toxicity of titanium dioxide P25 at repeated dose 28-day and 90-day in rats. Part Fibre Toxicol, 17, 34.
- Heringa MB, Peters RJB, Bleys R, van der Lee MK, Tromp PC, van Kesteren PCE, van Eijkeren JCH, Undas AK, Oomen AG and Bouwmeester H, 2018. Detection of titanium particles in human liver and spleen and possible health implications. Particle and Fibre Toxicology, 15, 15.
- Holder MK and Chassaing B, 2018. Impact of food additives on the gut-brain axis. Physiology & Behavior.
- Hong BY, Ideta T, Lemos BS, Igarashi Y, Tan Y, DiSiena M, Mo A, Birk JW, Forouhar F, Devers TJ, Weinstock GM and Rosenberg DW, 2019. Characterization of mucosal dysbiosis of early colonic neoplasia. NPJ Precis Oncol, 3, 29
- Hong J, Wang L, Zhao X, Yu X, Sheng L, Xu B, Liu D, Zhu Y, Long Y and Hong F, 2014. Th2 factors may be involved in TiO₂ NP-induced hepatic inflammation. Journal of Agriculture and Food Chemistry, 62, 6871–6878.
- Hong J, Hong FS, Ze YG and Zhang YQ, 2016. The nano-TiO₂ exposure can induce hepatic inflammation involving in a JAK-STAT signalling pathway. Journal of Nanoparticle Research, 18, 9.
- Hu R, Zheng L, Zhang T, Gao G, Cui Y, Cheng Z, Cheng J, Hong M, Tang M and Hong F, 2011. Molecular mechanism of hippocampal apoptosis of mice following exposure to titanium dioxide nanoparticles. Journal of Hazardous Materials, 191, 32–40.
- Hu H, Guo Q, Wang C, Ma X, He H, Oh Y, Feng Y, Wu Q and Gu N, 2015. Titanium dioxide nanoparticles increase plasma glucose via reactive oxygen species-induced insulin resistance in mice. Journal of Applied Toxicology: JAT, 35, 1122–1132.
- Huang S, Chueh PJ, Lin YW, Shih TS and Chuang SM, 2009. Disturbed mitotic progression and genome segregation are involved in cell transformation mediated by nano-TiO₂ long-term exposure. Toxicology and Applied Pharmacology, 241, 2.



- Hufnagel M, Schoch S, Wall J, Strauch BM and Hartwig A, 2020. Toxicity and gene expression profiling of copperand titanium-based nanoparticles using air-liquid interface exposure. Chemical Research in Toxicology, 33, 1237–1249.
- ISO/TR,, 2019. Nanotechnologies considerations for performing toxicokinetic studies with nanomaterials. ISO/TR 22019;2019, IDT.
- Ivett JL, Brown BM, Rodgers C, Anderson BE, Resnick MA and Zeiger E, 1989. Chromosomal aberrations and sister chromatid exchange tests in Chinese hamster ovary cells in vitro. IV. Results with 15 chemicals. Environmental and Molecular Mutagenesis, 14, 165–187.
- Jain AK, Senapati VA, Singh D, Dubey K, Maurya R and Pandey AK, 2017. Impact of anatase titanium dioxide nanoparticles on mutagenic and genotoxic response in Chinese hamster lung fibroblast cells (V-79): the role of cellular uptake. Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association, 105, 127–139.
- Jayaram DT and Payne CK, 2020. Intracellular generation of superoxide by TiO₂ nanoparticles decreases histone deacetylase 9 (HDAC9), an epigenetic modifier. Bioconjugate Chem, 31, 1354–1361.
- JECFA (Joint FAO, WHO Expert Committee on Food Additives), 2006. Titanium dioxide: Chemical and Technical Assessment. First draft prepared by P.M. Kuznesof, reviewed by M.V. Rao.
- Jensen DM, Lohr M, Sheykhzade M, Lykkesfeldt J, Wils RS, Loft S and Moller P, 2019. Telomere length and genotoxicity in the lung of rats following intragastric exposure to food-grade titanium dioxide and vegetable carbon particles. Mutagenesis, 34, 203–214.
- Jiang Z, Zhao M, Zhang H, Li Y, Liu M and Feng F, 2018. Antimicrobial emulsifier- glycerol monolaurate induces metabolic syndrome, gut microbiota dysbiosis and systemic low-grade inflammation in low-fat diet fed mice. Molecular Nutrition and Food Research, 1700547, 1–11.
- Jin C, Tang Y, Fan XY, Ye XT, Li XL, Tang K, Zhang YF, Li AG and Yang YJ, 2013. In vivo evaluation of the interaction between titanium dioxide nanoparticle and rat liver DNA. Toxicology and Industrial Health, 29.
- Jugan M-L, Barillet S, Simon-Deckers A, Herlin-Boime N, Sauvaigo S, Douki T and Carriere M, 2012. Titanium dioxide nanoparticles exhibit genotoxicity and impair DNA repair activity in A549 cells. Nanotoxicology, 6, 501–513.
- Kada T, Hirano K and Shirasu Y, 1980. Screening of environmental chemical mutagens by the rec-assay system with Bacillus subtilis. In: de Serres FJ and Hollaender A (eds.). Chemical Mutagens. Principles and Methods for their Detection. Vol 6. Plenum Press, New York, NY. pp. 149–173.
- Kandeil MA, Mohammed ET, Hashem KS, Aleya L and Abdel-Daim MM, 2019. Moringa seed extract alleviates titanium oxide nanoparticles (TiO₂-NPs)-induced cerebral oxidative damage, and increases cerebral mitochondrial viability. Environmental Science and Pollution Research International.
- Kang SJ, Kim BM, Lee YJ and Chung HW, 2008. Titanium dioxide nanoparticles trigger p53-mediated damage response in peripheral blood lymphocytes. Environmental and Molecular Mutagenesis, 49, 399–405.
- Karia B, Martinez JM and Bishop AJR, 2013. Induction of homologous recombination following in utero exposure to DNA-damaging agents. DNA Repair (Amst), 12, 912–921.
- Karimi S, Khorsandi L and Nejaddehbashi F, 2019. Protective effects of Curcumin on testicular toxicity induced by titanium dioxide nanoparticles in mice. JBRA Assisted Reproduction, 23, 344–351.
- Karimipour M, Zirak JM, Ahmadi A and Jafari A, 2018. Oral administration of titanium dioxide nanoparticle through ovarian tissue alterations impairs mice embryonic development. International Journal of Reproductive Biomedicine (Yazd, Iran), 16, 397–404.
- Karlsson HL, Gustafsson J, Cronholm P and M€oller L, 2009. Size-dependent toxicity of metal oxide particles a comparison between nano- and micrometer size. Toxicology Letters, 188, 112–118.
- Kathawala MH, Yun Z, Chu JJH, Ng KW and Loo SCJ, 2015. TiO_2 -nanoparticles shield HPEKs against ZnO-induced genotoxicity. Materials & Design, 88, 41–50.
- Kazimirova A, Baranokova M, Staruchova M, Drlickova M, Volkovova K and Dusinska M, 2019. Titanium dioxide nanoparticles tested for genotoxicity with the comet and micronucleus assays in vitro, ex vivo and in vivo. Mutation Research, 843, 57–65.
- Kazimirova A, El Yamani N, Rubio L, García-Rodríguez A, Barancokova M, Marcos R and Dusinska M, 2020. Effects of titanium dioxide nanoparticles on the hprt gene mutations in V79 hamster cells. Nanomaterials, Basel, 10.
- Khan M, Naqvi AH and Ahmad M, 2015. Comparative study of the cytotoxic and genotoxic potentials of zinc oxide and titanium dioxide nanoparticles. Toxicology Reports, 2, 765–774.
- Khorsandi L, Orazizadeh M, Mansouri E, Hemadi M and Moradi-Gharibvand N, 2016. Morphometric and stereological assessment of the effects of titanium dioxide nanoparticles on the mouse testicular tissue. Bratislavske lekarske listy, 117, 659–664.
- Khorsandi L, Orazizadeh M, Moradi-Gharibvand N, Hemadi M and Mansouri E, 2017. Beneficial effects of quercetin on titanium dioxide nanoparticles induced spermatogenesis defects in mice. Environmental Science and Pollution Research International, 24, 5595–5606.
- Kim JA, Aberg C and Dawson KA, 2012. Role of cell cycle on the cellular uptake and dilution of nanoparticles in a cell population. Nature Nanotechnology, 7, 62–68.



- Kim N, Kim C, Jung S, Park Y, Lee Y, Jo J, Hong M, Lee S, Oh Y and Jung K, 2018. Determination and identification of titanium dioxide nanoparticles in confectionery foods, marketed in South Korea, using inductively coupled plasma optical emission spectrometry and transmission electron microscopy. Food Additives & Contaminants. Part A, Chemistry, Analysis, Control, Exposure & Risk Assessment, 35, 1238–1246.
- Klawonn X, et al., 2017 a-f. Study summaries reported in the registration dossier under the REACH Regulation. Available online: https://echa.europa.eu/es/registration-dossier/-/registered-dossier/15560/4/9
- Klimisch HJ, Andreae M and Tillmann U, 1997. A systematic approach for evaluating the quality of experimental toxicological and ecotoxicological data. Regulatory Toxicology and Pharmacology, 25, 1–5.
- Knaapen AM, Borm PJA, Albrecht C and Schins RPF, 2004. Inhaled particles and lung cancer. Part A. Mechanisms. International Journal of Cancer, 109, 799–809.
- Kowalczyk M, Orłowski M, Klepacki Ł, Zinkiewicz K, Kurpiewski W, Kaczerska D, Pesta W, Zieliński E and Siermontowski P, 2020. Rectal aberrant crypt foci (ACF) as a predictor of benign and malignant neoplastic lesions in the large intestine. BMC Cancer, 20, 133.
- Kreyling WG, Holzwarth U, Haberl N, Kozempel J, Hirn S, Wenk A, Schleh C, Schaffler M, Lipka J, Semmler-Behnke M and Gibson N, 2017a. Quantitative biokinetics of titanium dioxide nanoparticles after intravenous injection in rats: part 1. Nanotoxicology., 11, 434–442.
- Kreyling WG, Holzwarth U, Schleh C, Kozempel J, Wenk A, Haberl N, Hirn S, Schaffler M, Lipka J, Semmler-Behnke M and Gibson N, 2017b. Quantitative biokinetics of titanium dioxide nanoparticles after oral application in rats: part 2. Nanotoxicology, 11, 443–453.
- Kumar S, Meena R and Paulraj R, 2016. Role of macrophage (M1 and M2) in titanium-dioxide nanoparticle-induced oxidative stress and inflammatory response in rat. Applied Biochemistry and Biotechnology, 180, 1257–1275.
- Kumar S, Hussain A, Bhushan B and Kaul G, 2020. Comparative toxicity assessment of nano- and bulk-phase titanium dioxide particles on the human mammary gland in vitro. Human and Experimental Toxicology, 39, 1475–1486
- Kurzawa-Zegota M, Sharma V, Najafzadeh M, Reynolds PD, Davies JP, Shukla RK, Dhawan A and Anderson D, 2017. Titanium dioxide nanoparticles induce DNA damage in peripheral blood lymphocytes from polyposis coli, colon cancer patients and healthy individuals: an ex vivo/in vitro study. Journal of Nanoscience and Nanotechnology, 17, 9274–9285.
- Landsiedel R, Fabian E, Ma-Hock L, van Ravenzwaay B, Wohlleben W, Wiench K and Oesch F, 2012. Toxico-/biokinetics of nanomaterials. Archives of Toxicology, 86, 1021–1060.
- Lazic V, Vukoje I, Milicevic B, Spremo-Potparevic B, Zivkovic L, Topalovic D, Bajic V, Sredojevic D and Nedeljkovic JM, 2019. Efficiency of the interfacial charge transfer complex between TiO₂ nanoparticles and caffeic acid against DNA damage in vitro: a combinatorial analysis. Journal of the Serbian Chemical Society, 84, 539–553.
- Lecomte M, Couedelo L, Meugnier E, Plaisancie P, Letisse M, Berengere B, Gabert L, Penhoat A, Durand A, Pineau G, Joffre F, Géloën A, Vaysse C, Laugerette F and Michalski MC, 2016. Dietary emulsifiers from milk and soybean differently impact adiposity and inflammation in association with modulation of colonic goblet in high fat fed mice. Molecular Nutrition and Food Research, 60, 606–620.
- Lee J, Jeong JS, Kim SY, Park MK, Choi SD, Kim UJ, Park K, Jeong EJ, Nam SY and Yu WJ, 2019. Titanium dioxide nanoparticles oral exposure to pregnant rats and its distribution. Particle and Fibre Toxicology, 16, 31.
- Li N, Ma L and Wang J, 2010. Interaction Between Nano-Anatase TiO_2 and Liver DNA from Mice In Vivo. Nanoscale Research Letters, 108.
- Li Y, Doak SH, Yan J, Chen DH, Zhou M, Mittelstaedt RA, Chen Y, Li C and Chen T, 2017a. Factors affecting the in vitro micronucleus assay for evaluation of nanomaterials. Mutagenesis, 32, 151–159.
- Li Y, Yan J, Ding W, Chen Y, Pack LM and Chen T, 2017b. Genotoxicity and gene expression analyses of liver and lung tissues of mice treated with titanium dioxide nanoparticles. Mutagenesis, 32, 33–46.
- Li J, Yang S, Lei R, Gu W, Qin Y, Ma S, Chen K, Chang Y, Bai X, Xia S, Wu C and Xing G, 2018. Oral administration of rutile and anatase TiO₂ nanoparticles shifts mouse gut microbiota structure. Nanoscale, 10, 7736–7745.
- Li XB, Zhang YS, Li B, Cui J, Gao N, Sun H, Meng QT, Wu SS, Bo JZ, Yan LC, Wu J and Chen R, 2019. Prebiotic protects against anatase titanium dioxide nanoparticles-induced microbiota-mediated colonic barrier defects. Nanoimpact, 14, 9.
- Liao F, Chen L, Liu Y, Zhao D, Peng W, Wang W and Feng S, 2019. The size-dependent genotoxic potentials of titanium dioxide nanoparticles to endothelial cells. Environmental Toxicology, 34, 1199–1207.
- Lim JH, Bae D and Fong A, 2018. Titanium dioxide in food products: quantitative analysis using icpms and raman spectroscopy. Journal of Agricultural and Food Chemistry, 66, 13533–13540.
- Linnainmaa K, Kivipensas P and Vainio H, 1997. Toxicity and cytogenetic studies of ultrafine titanium dioxide in cultured rat liver epithelial cells. Toxicology In Vitro, 1997, 329–335.
- Lomer MC, Thompson RP, Commisso J, Keen CL and Powell JJ, 2000. Determination of titanium dioxide in foods using inductively coupled plasma optical emission spectrometry. Analyst, 125, 2339–2343.
- Lotfi A, Zirak RG, Moghadam MS and Pazooki N, 2016. Effects of the interaction of nanorutile TiO_2 with vincristine sulfate on chromosomal abnormalities in vivo. International Journal of Advanced Biotechnology and Research, 7, 1083–1093.



- Louro H, Tavares A, Vital N, Costa PM, Alverca E, Zwart E, de Jong W, Fessard V, Lavinha J and Silva MJ, 2014. Integrated approach to the in vivo genotoxic effects of a titanium dioxide nanomaterial using LacZ plasmid based transgenic mice. Environmental and Molecular Mutagenesis, 55, 500–509. https://doi.org/10.1002/em. 21864
- Lu P-J, Ho I-C and Lee T-C, 1998. Induction of sister chromatid exchanges and micronuclei by titanium dioxide in Chinese hamster ovary-K1 cells. Mutation Research/Genetic Toxicology and Environmental Mutagenesis, 414, 15–20.
- Lu T, Ling C, Hu M, Meng X, Deng Y, An H, Li L, Hu Y, Wang H, Song G and Guo S, 2020. Effect of nano-titanium dioxide on blood-testis barrier and MAPK signaling pathway in male mice. Biological Trace Element Research.
- Ma Y, Guo YS, Wu S, Lv ZQ, Zhang Q and Ke YB, 2017. Titanium dioxide nanoparticles induce size-dependent cytotoxicity and genomic DNA hypomethylation in human respiratory cells. Rsc Advances, 7, 23560–23572.
- Magdolenova Z, Bilani D, Pojana G, Fjellsbø LM, Hudecova A, Hasplova K, Marcomini A and Dusinska M, 2012. Impact of agglomeration and different dispersions of titanium dioxide nanoparticles on the human related in vitro cytotoxicity and genotoxicity. Journal of Environmental Monitoring, 14, 455.
- Manivannan J, Banerjee R and Mukherjee A, 2020. Genotoxicity analysis of rutile titanium dioxide nanoparticles in mice after 28 days of repeated oral administration. Nucleus-India, 63, 17–24.
- Marion-Letellier R, Amamou A, Savoye G and Ghosh S, 2019. Inflammatory bowel diseases and food additives: to add fuel on the flames!. Nutrients, 11, 1–12.
- Martins Jr ADC, Azevedo LF, de Souza Rocha CC, Carneiro MFH, Venancio VP, de Almeida MR, Antunes LMG, de Carvalho Hott R, Rodrigues JL, Ogunjimi AT, Adeyemi JA and Barbosa Jr F, 2017. Evaluation of distribution, redox parameters, and genotoxicity in Wistar rats co-exposed to silver and titanium dioxide nanoparticles. Journal of Toxicology and Environmental Health. Part A, 80, 1156–1165.
- Meacock G,Taylor KDA, Knowles MJ and Himonides A, 1997. The improved whitening of minced cod flesh using dispersed titanium dioxide. Journal of the Science of Food and Agriculture, 73, 221E225
- Meena R, Kajal K and Paluraj R, 2015a. Cytotoxic and genotoxic effects of titanium dioxide nanoparticles in testicular cells of male Wistar rat. Applied Biochemistry and Biotechnology, 175, 825–840.
- Meena R, Kumar S and Paulraj R, 2015b. Titanium oxide (TiO₂) nanoparticles in induction of apoptosis and inflammatory response in brain. Journal of Nanoparticle Research, 17, 14.
- Merga Y, Campbell BJ and Rhodes JM, 2014. Mucosal barrier, bacteria and inflammatory bowel disease: possibilities for therapy. Digestive Diseases, 32, 475–483.
- Miller BM, Pujadas E and Gocke E, 1995. Evaluation of the micronucleus test in vitro using Chinese hamster cells: results of four chemicals weakly positive in, in vivo micronucleus test. Environmental and Molecular Mutagenesis, 26, 240–247.
- Minigalieva IA, Katsnelson BA, Privalova LI, Sutunkova MP, Gurvich VB, Shur VY, Shishkina EV, Valamina IE, Makeyev OH, Panov VG, Varaksin AN, Bushueva TV, Sakhautdinova RR, Klinova SV, Solovyeva SN and Meshtcheryakova EY, 2018. Combined Subchronic Toxicity of Aluminum (III), Titanium (IV) and Silicon (IV) Oxide Nanoparticles and Its Alleviation with a Complex of Bioprotectors. International Journal of Molecular Sciences, 19, 28.
- Modrzynska J, Berthing T, Ravn-Haren G, Jacobsen NR, Weydahl IK, Loeschner K, Mortensen A, Saber AT and Vogel U, 2018. Primary genotoxicity in the liver following pulmonary exposure to carbon black nanoparticles in mice. Particle and Fibre Toxicology, 15, 2.
- Mohamed HR, 2015. Estimation of TiO_2 nanoparticle-induced genotoxicity persistence and possible chronic gastritis-induction in mice. Food and Chemical Toxicology: AN International Journal Published for the British Industrial Biological Research Association, 83, 76–83.
- Mohamed HR and Hussien NA, 2016. Genotoxicity studies of titanium dioxide nanoparticles (TiO₂NPs) in the brain of mice. Scientifica, 2016, 6710840.
- Mohammadipour A, Hosseini M, Fazel A, Haghir H, Rafatpanah H, Pourganji M and Bideskan AE, 2016. The effects of exposure to titanium dioxide nanoparticles during lactation period on learning and memory of rat offspring. Toxicology and Industrial Health, 32, 221–228.
- Mohamed Magdy Badr El Dine F, Samie Mohamed Hussein HA and Abdelmawla Ghazala R, 2018. Evaluation of epigenetic changes of liver tissue induced by oral administration of Titanium dioxide nanoparticles and possible protective role of Nigella Sativa oil, in adult male albino rats. Nanomedicine Journal, 5, 192–198.
- Møller P, Jantzen K, Løhr M, Andersen MH, Jensen DM, Roursgaard M, Danielsen PH, Jensen A and Loft S, 2018. Searching for assay controls for the Fpg- and hOGG1-modified comet assay. Mutagenesis, 33, 9–19. https://doi.org/10.1093/mutage/gex015.
- Moran-Martinez J, del Rio-Parra RB, Betancourt-Martinez ND, Garcia-Garza R, Jimenez-Villarreal J, Nino-Castaneda MS, Nava-Rivera LE, Umana JAF, Carranza-Rosales P and Perez-Vertti RDA, 2018. Evaluation of the coating with TiO₂ nanoparticles as an option for the improvement of the characteristics of NiTi archwires: histopathological, cytotoxic, and genotoxic evidence. Journal of Nanomaterials, 11.
- Mottola F, Iovine C, Santonastaso M, Romeo ML, Pacifico S, Cobellis L and Rocco L, 2019. NPs-TiO₂ and Lincomycin Coexposure Induces DNA Damage in Cultured Human Amniotic Cells. Nanomaterials (Basel, Switzerland), 9 pp.



- Murugadoss S, Brassinne F, Sebaihi N, Petry J, Cokic SM, Van Landuyt KL, Godderis L, Mast J, Lison D, Hoet PH and van den Brule S, 2020. Agglomeration of titanium dioxide nanoparticles increases toxicological responses in vitro and in vivo. Part Fibre Toxicol, 17, 10.
- Myhr BC and Caspary WJ, 1991. Chemical mutagenesis at the thymidine kinase locus in L5178Y mouse lymphoma cells: results for 31 coded compounds in the National Toxicology Program. Environmental and Molecular Mutagenesis, 18, 51–83.
- Nakagawa Y, Wakuri S, Sakamoto K and Tanaka N, 1997. The photogenotoxicity of titanium dioxide particles. Mutation Research, 394, 125–132.
- Naya M, Kobayashi N, Ema M, Kasamoto S, Fukumuro M, Takami S, Nakajima M, Hayashi M and Nakanishi J, 2012. In vivo genotoxicity study of titanium dioxide nanoparticles using comet assay following intratracheal instillation in rats. Regulatory Toxicology and Pharmacology, 62, 1–6.
- NCI, 1979. National Toxicology Program. Bioassay of titanium dioxide for possible carcinogenicity. Natl Cancer Inst Carcinog Tech Rep Ser., 97, 1–123.
- Nejrup RG, Licht TR and Hellgren LI, 2017. Fatty acid composition and phospholipid types used in infant formulas modifies the establishment of human gut bacteria in germ-free mice. Scientific Reports, 7, 1–11.
- Niu L, Shao M, Liu Y, Hu J, Li R, Xie H, Zhou L, Shi L, Zhang R and Niu Y, 2017. Reduction of oxidative damages induced by titanium dioxide nanoparticles correlates with induction of the Nrf2 pathway by GSPE supplementation in mice. Chemico-Biological Interactions, 275, 133–144.
- NVWA (Netherlands Food and Consumer Product Safety Authority), 2019. Opinion of BuRO on possible health effects of the food additive titanium dioxide (E171). Trcvwa/2019/4476/EN.
- OECD (Organisation for Economic Co-operation and Development), 2005. Manual for the investigation of HPV chemicals. Chapter 3.1 Guidance for Determining the Quality of Data for the SIDS Dossier (Reliability, Relevance and Adequacy). Available online: http://www.oecd.org/chemicalsafety/risk-assessment/49191960.pdf
- OECD (Organisation for Economic Co-operation and Development), 2014. Report on statistical issues related to OECD Test Guidelines (TGs) on genotoxicity, ENV/JM/MONO(2014)12.
- OECD, 2016. Environment directorate joint meeting of the chemicals committee and the working party on chemicals, pesticides and biotechnology. Titanium dioxide: summary of the dossier. Vol No. 73. (Organisation for Economic Co-operation and Development). ENV/JM/MONO(2016) 25.
- Olmedo DG, Tasat DR, Evelson P, Guglielmotti MB and Cabrini Cabrini RL, 2008. Biological response of tissues with macrophagic activity to titanium dioxide. Journal of Biomedical Materials Research, 84A, 1087.
- Osman IF, Baumgartner A, Cemeli E, Fletcher JN and Anderson D, 2010. Genotoxicity and cytotoxicity of zinc oxide and titanium dioxide in HEp-2 cells. Nanomedicine, 5, 1193–1203.
- Osman IF, Najafzadeh M, Sharma V, Shukla RK, Jacob BK, Dhawan A and Anderson D, 2018. TiO₂ NPs induce DNA damage in lymphocytes from healthy individuals and patients with respiratory diseases-an ex vivo/in vitro study. Journal of Nanoscience and Nanotechnology, 18, 544–555.
- Panté N and Kann M, 2002. Nuclear pore complex is able to transport macromolecules with diameters of about 39 nm. Molecular Biology of the Cell, 13, 425–434.
- Patel S, Patel P, Sachin B, Undre SR, Pandya MS and Sonal B, 2016. DNA binding and dispersion activities of titanium dioxide nanoparticles with UV/vis spectrophotometry, fluorescence spectroscopy and physicochemical analysis at physiological temperature. Journal of Molecular Liquids, 213.
- Patel S, Patel P and Bakshi SR, 2017. Titanium dioxide nanoparticles: an in vitro study of DNA binding, chromosome aberration assay, and comet assay. Cytotechnology, 69, 245–263.
- Pele LC, Thoree V, Bruggraber S, Koller D, Thompson RP, Lomer MC and Powell JJ, 2015. Pharmaceutical/food grade titanium dioxide particles are absorbed into the bloodstream of human volunteers. Particle and Fibre Toxicology, 12, 26.
- Peters RJB, Van Bemmel G, Herrera-Rivera Z, Helsper HPFG, Marvin HJP, Weigel S, Tromp PC, Oomen AG, Rietveld AG and Bouwmeester H, 2014. Characterization of titanium dioxide nanoparticles in food products: analytical methods to define nanoparticles. Journal of Agricultural and Food Chemistry, 62, 6285–6293.
- Peters RJB, Oomen AG, van Bemmel G, van Vliet L, Undas AK, Munniks S, Bleys RLAW, Tromp PC, Brand W and van der Lee M, 2020. Silicon dioxide and titanium dioxide particles found in human tissues. Nanotoxicology, 14, 420–432.
- Phillips LG and Barbano DM, 1997. The influence of fat substitutes based on protein and titanium dioxide on the sensory properties of lowfat milks. Journal of Dairy Sciences, 80, 2726.
- Pichot R, Duffus L, Zafeiri I, Spyropoulos F and Norton IT, 2015. Particle-Stabilized Food Emulsions. Particle-Stabilized Emulsions and Colloids. Formation and Applications, RSC.
- Pinget G, Tan J, Janac B, Kaakoush NO, Angelatos AS, O'Sullivan J, Koay YC, Sierro F, Davis J, Divakarla SK, Khanal D, Moore RJ, Stanley D, Chrzanowski W and Macia L, 2019. Impact of the food additive titanium dioxide (E171) on gut microbiota-host interaction. Frontiers in Nutrition, 6, 57.
- Pittol M, Tomacheski D, Simoes DN, Ribeiro VF and Santana RMC, 2018. Evaluation of the toxicity of silver/silica and titanium dioxide particles in mammalian cells. Brazilian Archives of Biology and Technology, 61, 14.
- Pogribna M, Koonce NA, Mathew A, Word B, Patri AK, Lyn-Cook B and Hammons G, 2020. Effect of titanium dioxide nanoparticles on DNA methylation in multiple human cell lines. Nanotoxicology, 1–20.